

## AUSTRALIAN NEREIDAE

Including descriptions of three new species and one genus,  
together with summaries of previous records and keys to species.

By OLGA HARTMAN\*

Communicated by S. J. Edmonds

[Read 8 May 1953]

### SUMMARY

The Australian (and New Zealand) Nereidae are recorded with 47 species in 13 genera. One genus, *Anstralonereis* is new; two species, *Ceratocephala edmondsi* and *Micronereis halei*, and one subspecies, *Platynereis dianeriki antipoda* are newly described. There are many new records of distribution, particularly for the species occurring in the Flindersian and Peronian provinces. The recorded data of all the species are summarised in a series of charts.

### INTRODUCTION

The polychaetous annelids of the family Nereidae are among the more conspicuous, well represented groups of marine invertebrates in the Commonwealth of Australia and the Dominion of New Zealand. As in other known geographic areas, they are largely littoral. Curiously, however, the present study indicates that the nereids, at least for the southern half of Australia, are unusually diversified and modified, probably more so than in any other geographic area of comparable size. Thus, among the 47 species in 13 genera, there are some with very primitive characters, such as presence of setae in the first segment; others, such as *Anstralonereis*, have functional coelomoducts and papillated ventrum. These facts make it particularly desirable to recognize their positions or affinities with the nereids of other parts of the world.

In spite of the fairly large number (47) recorded here, it can hardly be assumed that the number of species is even nearly complete. Much of the coastline remains almost unknown with respect to its polychaetous fauna. The records to date are largely those made by incidental collecting. There have been no extensive surveys of coastal areas such as was done for the echinoderms (Clark, 1946).

Recent studies by Knox (1951) on the nereids of New Zealand indicate that there are conspicuous differences in the fauna of the Dominion and the Commonwealth, at least for its southern half. Comparison has been difficult in many cases for the literature is scattered and sometimes obscure in essential details. Type collections, if existing, are often deposited in museums outside of Australia.

An attempt is here made to correlate and assemble these scattered data. Charts I to IV summarize the records of the 47 species, including: acceptable name, date and source of original publication, place of origin, diagnostic accounts, synonyms, distributional data and new records, ecologic niche, unique characteristics, method of reproduction in so far as known, and the formulae of the proboscoidal processes.

The materials on which these studies are based were collected mainly from littoral zones of South Australia, Victoria and New South Wales, thus are largely south-eastern Australia. These areas fall within the Peronian and Flindersian provinces of Hedley. Based on studies of the echinoderm fauna, H. L. Clark (1946) finds that these two provinces have the most numerous endemic species (82% of the Peronian and 89% of the Flindersian echinoderms are endemic in Australia). If the annelids are equally unique, as a comparison of the charts indicates, one may expect a widely diversified polychaetous fauna.

\* Allan Hancock Foundation of the University of Southern California.

Contribution No. 138.

Trans. Roy Soc. S. Aust., 77, July, 1954

I am indebted to the following people and institutions for the collections on which the present study is based: Mr. Herbert M. Hale and associates of the South Australian Museum, Mr. S. J. Edmonds of the University of Adelaide, and Miss Barbara Dew of Cronulla, New South Wales. The Administration of the Allan Hancock Foundation of the University of Southern California provided material aid and support to conduct these studies. Illustrations are by Anker Petersen of the Allan Hancock Foundation.

Types and complete series of species are deposited in the institutions from which the collections originate. Duplicate series are in the Allan Hancock Foundation.

CHART I.

Name of Species	Date and source of publication	Place of origin	Diagnostic account	Synonyms
<i>Australonereis ehlersi</i> (Augener)	1913, pp. 142-145, figs.	Swan River area, W.A.	Monro, 1938, pp. 618-623, and below	<i>Nereis</i> ( <i>Leonnates</i> ) <i>ehlersi</i> Augener, 1913, and <i>Leptonereis ehlersi</i> Monro, 1938
<i>Ceratocephala edmondssi</i> , n. sp.	Herein	South Australia	Below	—
<i>Ceratonereis mirabilis</i> Kinberg	1866, p. 170	Brazil	Augener, 1913, pp. 168-171; Horst, 1924, pp. 180-182 figs.	<i>Ceratonereis tentaculata</i> Kinberg, 1866
<i>Ceratonereis lapinigenis</i> Grube	1878, pp. 69-70	Philippine Islands	Augener, 1913, pp. 166-168	Possibly the same as <i>C. costae</i> Grube, 1840
<i>Ceratonereis aequisetis</i> Augener	1913, pp. 171-174 figs.	Swan River area W.A.	(See original)	Questionably includes <i>C. erythraeensis</i> (fide Monro, 1938)
<i>Ceratonereis erythraeensis</i> Fauvel	1918, pp. 505-506 figs.	Madagascar	Fauvel, 1919, pp. 407-410 figs.	Possibly the same as <i>S. aequisetis</i> Augener
<i>Cheltonereis peristomialis</i> Benham	1916a, pp. 392-393	New Zealand and South Australia	Benham, 1916b, pp. 138-143, figs.	—
<i>Micronereis halei</i> , n. sp.	Herein	South Australia	Below	—
<i>Nannanereis* quadraticeps</i> (Blanchard)	1849, p. 25, figs.	Chile	Benham, 1909, pp. 242-244, figs.	<i>Lycastis quadraticeps</i> Blanchard 1849 and others
<i>Nicon aestuariensis</i> Knox	1951, pp. 225-227, figs.	Banks Peninsula, N.Z.	(See original)	—
<i>Neanthes vaalii</i> Kinberg	1866, p. 171	Pt. Jackson, N.S.W.	Augener, 1913, pp. 149-153; Augener, 1924, p. 316	<i>Nereis albanensis</i> Augener
<i>Neanthes cricognatha</i> (Ehlers)	1905, pp. 29-30 figs.	New Zealand	Benham, 1916, p. 134	<i>Nereis arenaceodentata</i> Benham, 1916
<i>Neanthes</i> , near <i>cricognatha</i> (Ehlers)	Herein	South Australia	Below	—
<i>Neanthes kerguelensis</i> (McIntoshi)	1885, pp. 225-227 figs.	Kerguelen Islands	Augener, 1924, p. 330; Augener, 1913, pp. 164-166	<i>Nereis kerguelensis</i> var. <i>oligodonta</i> Augener, 1913 (?) (see below)
<i>Neanthes orypoda</i> (Marenzeller)	1879, pp. 120-121 figs.	Southern Japan	Monro, 1938, p. 614, figs.	<i>Nereis (Alitta) orypoda</i> Marenzeller, 1879

\* *Nannanereis* Chamberlin, 1919, has priority over *Lycastella* Feuerborn, 1931. Both generic names were proposed for the same species, *Lycastis quadraticeps* Blanchard, as type.

Name of Species	Date and source of publication	Place of origin	Diagnostic account	Synonyms
<i>Neanthes unifasciata</i> (Willey)	1905, pp. 271-277 figs.	Ceylon	Monro, 1931, p. 13	<i>Nereis unifasciata</i> Willey, 1905
<i>Neanthes angusticollis</i> (Augener) (not <i>Nereis angusticollis</i> Kbg.)	1913, pp. 145-149 figs.	Sharks Bay, W.A.	(See original)	—
<i>Eunereis marri</i> Monro	1939, p. 117 figs.	Off South-western Australia	(See original)	—
<i>Nereis falcata</i> (Willey)	1905, p. 117, figs.	Ceylon	Fauvel, 1921, pp. 8-11, figs. Augener, 1923, pp. 21-26, figs.	<i>Nereis kauderni</i> Fauvel, 1921 <i>Nereis mortenseni</i> Augener, 1923
<i>Nereis denhamensis</i> Augener	1913, pp. 156-159, figs.	Sharks Bay, W.A.	(See original and below)	—
<i>Nereis jacksoni</i> Kinberg	1866, p. 169	Pt. Jackson, N.S.W.	Augener, 1913, p. 159, figs. Augener, 1924, p. 317	<i>Nereis heinssonensis</i> Augener, 1913 p. 159, figs.
<i>Nereis thompsoni</i> Kott	1951, p. 103-105, figs.	Western Australia	Kott, 1951, p. 103-105, figs.	<i>Nereis (Neanthes) thompsoni</i> Kott, 1951
<i>Nereis peroniensis</i> Kott	1951, pp. 101-102, figs.	Point Peron, W.A.	(See original and below)	<i>Nereis callaena peroniensis</i> Kott, 1951
<i>Nereis cockburnensis</i> Augener	1913, pp. 153-156, figs.	Cockburn Sd., W.A.	(See original and below)	—
<i>Nereis languida</i> Kinberg	1866, p. 169	Pt. Jackson, N.S.W.	(See original; doubtful)	—
<i>Nereis robusta</i> Quatrefages	1865, pp. 544-545	New Zealand	(See original; doubtful)	—
<i>Perinereis amblyodonta</i> (Schmarda)	1861, p. 106 figs.	Pt. Jackson, N.S.W.	Knox, 1951, pp. 221-222, figs.	<i>Nereilepas amblyodonta</i> Schmarda, 1861; <i>Perinereis novae-hollandiae</i> Kinberg, 1866
<i>Perinereis variodontata</i> Augener	1913, pp. 179-182, figs.	Albany, W.A.	(See original and below)	—
<i>Perinereis vallata</i> (Grube)	1857, pp. 159-160	Chile	Knox, 1951, pp. 218-219, figs.	<i>Nereilepas pacifica</i> Schmarda, 1861
<i>Perinereis brevicirris</i> (Grube)	1866, pp. 176-177	St. Pauls Island ( <i>Noevara</i> Exp.)	Knox, 1951, pp. 219-220, figs.	<i>Perinereis heterodontata mictodon-</i> <i>toides</i> Augener, 1913
<i>Perinereis camiguinoides</i> Augener	1922a, pp. 180-183 figs.	Chile	Knox, 1951, pp. 220-221, figs.	<i>Nereis vancaurica</i> Ehlers, 1905
<i>Perinereis barbara</i> Monro	1926, pp. 318-320, figs.	N.S.W. and Eastern Australia	(See original)	—
<i>Perinereis pontiensis</i> Augener	1924, pp. 349-352, figs.	New Zealand	(See original)	—

Name of Species	Date and source of publication	Place of origin	Diagnostic account	Synonyms
<i>Perinereis pseudocamiguina</i> Augener	1922, pp. 183-186, figs.	Chile	Monro, 1939a, p. 118	Possibly the same as <i>Perinereis helleri</i> or <i>P. camiguina</i> (fide Monro 1939)
<i>Perinereis colmani</i> Monro	1926, pp. 318-320, figs.	N.S.W. and Eastern Australia	(See original)	—
<i>Perinereis nigropunctata</i> (Horst)	1889, pp. 171-174, figs.	Malay	Augener, 1929h, p. 24, figs.	<i>Perinereis yorkensis</i> Augener, 1922b
<i>Perinereis vancouverica</i> (Ehlers)	1868, p. XX	Nicobar Island	Monro, 1931, p. 14; Fauvel, 1933, pp. 34-35	<i>Nereis languida</i> Grube, 1868; <i>Perinereis nancaurica</i> (variant)
<i>Perinereis obfuscata</i> Grube	1878, pp. 86-87	Philippine Islands	Monro, 1931, pp. 16-18, figs.	—
<i>Perinereis helleri</i> Grube	1878, pp. 81-82	Philippine Islands	Monro, 1931, pp. 14-15, figs.	—
<i>Perinereis camiguina</i> Grube	1878, pp. 87-89, figs.	Philippine Islands	Monro, 1931, pp. 15-17, figs.	—
<i>Platynereis australis</i> (Schmarda)	1861, p. 101	New Zealand	Knox, 1951 (pp. 223-225, figs.)	<i>Heteronereis australis</i> Schmarda
<i>Platynereis dumerilii antipoda</i> , new species	Hercin	South Australia	(See below)	(see below)
<i>Platynereis magalhaensis</i> Kinberg	1866, p. 177	Straits of Magellan, Chile	Monro, 1930, pp. 106-107, figs. Monro, 1936, pp. 137-138	(see below)
<i>Platynereis polyscalma</i> Chamberlain	1919, pp. 219-226, figs.	South Pacific Islands	Fauvel, 1932, p. 114; and original	—
<i>Platynereis bicanaliculata</i> (Baird)	1863, p. 109	Vancouver Island, Canada	(See below)	(see below)
<i>Pseudonereis rothnestiana</i> Augener	1913, pp. 184-187	Rothnest, Green Is., W.A.	Augener, 1913, pp. 184-187	—
<i>Pseudonereis anomala</i> Gravier	1901, pp. 191-197, figs.	Red Sea	Fauvel, 1922, p. 494; Kott, 1951, pp. 93-95, figs.	<i>Nereis nichollsi</i> Kott, 1951

In addition, *Nereis ruficeps* Ehlers (1905, pp. 24-25, pl. 3, figs. 10-15) has been described from Chatham Island off New Zealand. It appears to be a species of *Neanthes* since notopodia have only spinigerous setae. It differs from other species of *Neanthes* recorded above in that areas V and VI of the proboscis are bare; tentacular cirri are articulated and rather short.

(?) *Nereis neozelandica* Benham listed by Knox (1951, p. 213) is presumably a manuscript name only.

CHART II.

Name of species	New locality	More extensive distribution	Ecologic niche
<i>Australonereis ehlersi</i>	Victoria at Lakes Entrance	Swan River area, Western Australia	In sandy mud flats, occupying thin, U-shaped tubes (B. Dew)
<i>Ceratocephala edmondsi</i>	Kangaroo Island, Sth. Aus.	—	Common in muddy and sandy tidal flats: tube sandy (S. J. Edmonds)
<i>Ceratonereis mirabilis</i>	Spencer Gulf, S.A.; western shoal in 30 feet	Circumundane in warm seas	In rock and shell gravel, in mud and on reefs (Monro, 1931)
<i>Ceratonereis lapinigenis</i>	Port Willunga, S.A.; Lakes Entrance, Vic.; Pitwater, Broken Bay, N.S.W.	Western Australia (Augener, 1913; Monro, 1939b)	On piles, sponges and bryozoans; along shore to 40 fms. (Monro, 1939b)
<i>Ceratonereis acquisetis</i>	—	Western Australia at the Swan River area	Along the strand in marine conditions (Augener, 1913)
<i>Ceratonereis erythraeensis</i>	—	Madagascar, and Swan River, W.A. (Monro, 1938)	Same as for <i>C. acquisetis</i> (Monro, 1938); in coral debris (Okuda, 1940, p. 9)
<i>Cheilonereis peristomidis</i>	—	New Zealand and South Australian Bight	In gastropod shell with hermit crab, or free-living (Benham, 1916)
<i>Micronereis halei</i>	Sellick Beach, Sth. Aus.	—	At exposed, outer edge of reef (H. M. Hale)
<i>Nanonereis quadraticeps</i>	California	Auckland Island (Augener, 1923); New Zealand (Benham, 1909); Southwest Africa (Augener, 1918) New Zealand	Under stones in varying salinities of high intertidal zones (Augener, 1923)
<i>Nicon aestuariensis</i>	—	—	Burrows in mud of estuaries with up to 3 hours exposure; number up to 150 per sq.m. (Knox, 1951)
<i>Neanthes vaalii</i>	Kangaroo Island, Sth. Aus.; New South Wales	New Zealand; Tasmania (Monro, 1939a)	Among sand, rock and weed (Knott, 1951)
<i>Neanthes cricognatha</i>	Sellick Beach and Port Adelaide, S.A.; Cronulla, N.S.W.	New Zealand; Western Australia (Knox, 1951, and Augener, 1913)	Intertidal to 55 fms.
<i>Neanthes near cricognatha</i>	American River, Kangaroo Island, Sth. Aus.	—	In colonies of the serpulid, <i>Galeolaria</i> (S. J. Edmonds)
<i>Neanthes kerguelensis</i>	Port Willunga and Sellick, Sth. Aus.	New Zealand; Tasmania and Sub-Antarctic (Augener, 1924)	On reefs and rocks; in sand (Augener, 1913)
<i>Neanthes oxyroda</i>	—	Japan; China; South-west Australia (Monro, 1938)	Mouth of the Swan River, W.A. (Monro, 1938)
<i>Neanthes unifasciata</i>	—	Great Barrier Reef (Monro, 1931)	On reefs (Monro, 1931)
<i>Neanthes angusticollis</i>	—	Sharks Bay, Western Australia	(Not known)

Name of Species	New locality	More extensive distribution	Ecologic niche
<i>Eumereis marri</i>	—	Off Southwestern Australia in 62 metres	(Not known)
<i>Nereis falcata</i>	—	Ceylon; Madagascar; New Zealand; subtropical (Knox, 1951; Fauvel, 1935, p. 302)	In sandy loam in 45 fms; under littoral stones (Augener, 1923)
<i>Nereis denhamensis</i>	New South Wales	Southwest Australia and South Australia (Benham, 1916)	Intertidal to 11 metres (Augener, 1913)
<i>Nereis jacksoni</i>	Sellick Beach, Sth, Aus.; New South Wales	S. and S.W. Australia; New South Wales (Monro, 1937; Fauvel, 1947; Augener, 1924)	Sandy or rocky shores, shore to 14 metres; on fouling plate in estuary (B. Dew)
<i>Neries thompsoni</i>	—	Rottnest, Point Peron and Aldrich's Cove, Normalup, W.A.	(Not known)
<i>Nereis peroniensis</i>	—	Western Australia	Probably rocks (Kott, 1951)
<i>Nereis cockburnensis</i>	Kangaroo Island and Sellick Beach, S.A.; N.S.W.	Western Australia	Among algae and their holdfasts on calcareous rock platforms (S. J. Edmonds) and from stones in rocky pools (H. M. Hale)
<i>Nereis langwida</i>	—	Port Jackson, N.S.W.	(Not known)
<i>Nereis robusta</i>	—	New Zealand	(Not known)
<i>Perinereis amblyodonta</i>	American River, Kangaroo Is.; Port Willunga, S.A.;	Southern and Western Australia; New Zealand; Indo-Pacific; Knox, 1951; Fauvel, 1947	Under rocks and in mud flats; on piles and mooring chains and in clumps of <i>Galeolaria</i> (S. J. Edmonds and B. Dew)
<i>Perinereis variodentata</i>	Port Jackson, N.S.W.; Sellick Beach, S.A.;	Western Australia (Augener, 1913)	From stones in rock pools and in limestone reefs (H. M. Hale)
<i>Perinereis vallata</i>	Wynyard, N.W., Tasmania	New Zealand; Southern Australia (Knox, 1951)	Intertidal, low water to 3 hour exposure; in mud among stones; in branching, slime-lined burrows (Knox, 1951)
<i>Perinereis brevicirris</i>	—	New Zealand; Indo-Pacific; Western Australia (Knox, 1951)	In estuaries; burrows in sand and mud; associated with <i>P. vallata</i> (Knox, 1951)
<i>Perinereis camigninoides</i>	—	Southern Chile; New Zealand	Intertidal, under stones, to 35 fms. (Knox, 1951)
<i>Perinereis barbara</i>	—	Port Jackson, N.S.W.	(Not known)
<i>Perinereis ponuiensis</i>	—	New Zealand; Auckland Islands; North Cape (Augener, 1924)	Under stones along the coast (Augener, 1924)
<i>Perinereis pseudocamignina</i>	—	Southern Chile; New Zealand; Tasmania (Monro, 1939)	Between tide marks, lower Derwent River, Hobart, Tasmania (Monro, 1939a)

Name of Species	New locality	More extensive distribution	Ecologic niche
<i>Perinereis culmani</i>	Pt. Jackson, Syd. Harbour, Pitwater, Broken Bay and Athol Bight, N.S.W.	China Sea and Eastern Australia (Monro, 1926)	Among tubes of <i>Galeolaria</i> , under rocks in kelp holdfasts and on piles (B. Dew)
<i>Perinereis nigropunctata</i>	—	Indo-Pacific; North Australia (Augener, 1922); Great Barrier Reef (Monro, 1931)	Associated with <i>Perinereis camiguina</i> (Augener, 1922)
<i>Perinereis vancaurica</i>	—	Great Barrier Reef (Monro, 1931); Philippines and New Zealand (Fauvel, 1923)	(Not known)
<i>Perinereis obfuscata</i>	—	Philippines; Indo-Pacific; Great Barrier Reef (Monro, 1931)	In lagoon of Great Barrier Reef (Monro, 1931)
<i>Perinereis helleri</i>	—	Indo-Pacific (Okuda, 1940); Great Barrier Reef (Monro, 1931)	In coral reef (Monro, 1931)
<i>Perinereis camiguina</i>	—	East and North Australia (Augener, 1922); Great Barrier Reef (Monro, 1931)	Among coral rocks (Monro, 1931)
<i>Platynereis australis</i>	—	New Zealand (Ehlers, 1905; Augener, 1926; Knox, 1951)	Among shell fragments and sand, occupying tubes; intertidal to 50 fms. (Knox, 1951)
<i>Platynereis dumerilii antipoda</i>	South Australia and New South Wales	—	Among algae, on piles, on mooring buoys, on limestone reefs (see below)
<i>Platynereis magalhaensis</i>	—	Sub-Antarctic and Southern New Zealand (Augener, 1923)	Among algae and holdfasts (Hartman, 1952, in press)
<i>Platynereis polyscalma</i>	—	Indo-Pacific (Fauvel, 1931); Great Barrier Reef (Monro, 1931)	Epitokous stages in plankton
<i>Platynereis bicamaliculata</i>	New South Wales and California (see below)	Hawaii; Northeast Pacific to West- ern Canada	Among algaec and holdfasts
<i>Pseudonereis rothnestiana</i>	—	Western Australia (Augener, 1913); Japan (Okuda, 1938)	On shallow reefs (Augener, 1913)
<i>Pseudonereis anomala</i>	—	Western Australia (Fauvel, 1922 and Kott, 1951); Red Sea (Gravier, 1901); East Indies (Horst, 1924)	Under stones (Gravely, 1927); among rocks and weed (Kott, 1951)



CHART III.

Name of species	Unique characteristics	Diagnostic setae	Method of reproduction	Position of modification, if any
<i>Australonereis ehlersi</i>	Ventrum with transverse rows of papillae	Falciigers limited to neuropodia	Presumably direct	—
<i>Ceratocephala edmondsi</i>	Long papillae on both rings of proboscis	Falciigers limited to neuropodia	(Not known)	—
<i>Ceratonereis mirabilis</i>	Anterior end of prostomium widely divergent; dorsal cirri very long	Falciigers limited to neuropodia	Epitoky (Horst, 1924 p. 180)	Male at 18/19
<i>Ceratonereis lapinigenis</i>	Anterior margin of prostomium entire; dorsal cirri not long	Falciigers limited to neuropodia	Epitoky (Kott, 1951, p. 107)	At 14/15 (sex not stated)
<i>Ceratonereis aequisetis</i>	Dorsal and ventral cirri greatly reduced	Falciigers limited to neuropodia	(Not known)	—
<i>Ceratonereis erythraeensis</i>	Large simple falciigers in posterior segments	Falciigers both simple and composite, limited to neuropodia	(Not known)	—
<i>Cheilonereis peristomialis</i>	Peristomium prolonged on ventral side to partly envelop prostomium	Falciigers limited to neuropodia	Epitoky (Benham, 1916, p. 142)	Female at 26/27
<i>Micronereis halei</i>	First segment is parapodial and setigerous	All setae of one kind, homomorphic spinigers	(Not known)	—
<i>Namanereis quadriceps</i>	Parapodia uniramous throughout	Spinigers above and falciigers below	Direct	—
<i>Nicon aestuariensis</i>	Proboscis lacks parapagnaths; parapodia biramous	Long-appendaged heteromorphic falciigers in neuropodia	Epitoky (Knox, 1951)	Body with 3 regions: 20 anterior, a median, and 20-30 posterior segments
<i>Neanthes vaalii</i>	High paragnathal count; parapodial lobes moderately small	Falciigers limited to neuropodia	Epitoky	Male at 18/19; Female at 20/21; with 2 body regions

Name of Species	Unique characteristics	Diagnostic setae	Method of reproduction	Position of modification, if any
<i>Neanthes cricognatha</i>	Oral ring of proboscis with continuous ring of cones	Falcigers limited to neuropodia	(Not known)	—
<i>Neanthes near cricognatha</i>	Oral ring with one row of large and many rows of tiny cones	Falcigers limited to neuropodia	(Not known)	—
<i>Neanthes kerguelensis</i>	Notopodial lobe diminishes in posterior segments	Falcigers limited to neuropodia	perhaps direct. (See Benham, 1916, page 133)	—
<i>Neanthes oxyboda</i>	Notopodial lobes greatly enlarged on both sides of base of dorsal cirrus	With spinigers only; falcigers absent	(Not known)	—
<i>Neanthes unifasciata</i>	Area V lacks paragnathis; dorsum with broad brown bands	Falcigers limited to neuropodia	Epitoky (Horst 1924 pp. 153-154)	Male at 16/17; Female at 18/19
<i>Neanthes angusticollis</i>	Dorsal lobe of posterior notopodia is triangular with dorsal cirrus attached at middle	Falcigers limited to neuropodia	(Not known)	—
<i>Eumereis marri</i>	Maxillary ring of proboscis lacks paragnathis	Notopodia with falcigers in which appendage is 2- or 3-dentate	(Not known)	—
<i>Nereis falcaria</i>	Prostomium incised in front; oral ring bare or with few paragnathis	Notopodial falcigers with appendage coarsely 2- or 3-dentate	Epitoky (Augener, 1914, p. 327)	Body with 3 regions in male; parapodial changes at 13/14 and last 50 segments unmodified
<i>Nereis denhamensis</i>	Notopodial lobe little reduced	Notopodial falcigers coarsely 2- or 3-dentate	Epitoky (Kott, 1951)	Body with 2 regions; change at 14/15 (sex not stated)
<i>Nereis jacksoni</i>	Notopodial lobe diminishes rapidly in back; proboscis with few cones	Notopodial falcigers coarsely 2- or 3-dentate	Absent or present (?) (Kott, 1951, p. 97)	At 13/14 (sex not stated)
<i>Nereis thompsoni</i>	Paragnathal count higher than in <i>N. denhamensis</i>	Notopodial falcigers coarsely dentate	Direct	—

Name of species	Unique characteristics	Diagnostic setae	Method of reproduction	Position of modification if any
<i>Nereis peroniensis</i>	Area III with 3 cones in a transverse row; base of notopodial lobe prolonged	Short - appendaged homomorph falcigers present in posterior segments	(Not known)	—
<i>Nereis cockburnensis</i>	Notopodial falcigers of 2 kinds	Homomorph falcigers present in all notopodia	Epitoky in April or later	Female at 16/17
<i>Nereis languida</i>	Obscure	(Not known)	(Not known)	—
<i>Nereis robusta</i>	Obscure	(Not known)	(Not known)	—
<i>Perinereis amblyodontia</i>	Posterior notopodial lobe prolonged, rectangular	Falcigers limited to notopodia	Epitoky (Augener, 1924, p. 343)	Female at 16 to 24/25
<i>Perinereis variodontata</i>	Areas V and VI of proboscis (see Chart IV)	Falcigers limited to notopodia	(Not known)	—
<i>Perinereis vallata</i>	Area V and VI of proboscis (see Chart IV)	Falcigers limited to notopodia	Subepitokes recorded Fauvel, 1919, p. 418)	—
<i>Perinereis brevicirris</i>	Area V of proboscis (see Chart IV)	Falcigers limited to notopodia	Perhaps direct (Knox 1951, p. 220)	—
<i>Perinereis camiguinoides</i>	Area V and VI of proboscis (see Chart IV)	Falcigers limited to notopodia	Epitoky (Knox, 1951, p. 221)	At 16/17 (sex not stated)
<i>Perinereis barbara</i>	Area V and VI of proboscis (see Chart IV)	Falcigers limited to notopodia; all spinigers homomorph	(Not known)	—
<i>Perinereis pomianensis</i>	Area V and VI of proboscis (see Chart IV)	Falcigers limited to notopodia; acicula occur by twos	(Not known)	—
<i>Perinereis pseudocamiguina</i>	Posterior notopodial lobe greatly prolonged with dorsal cirrus inserted near distal end	Falcigers limited to notopodia	(Not known)	—
<i>Perinereis calmani</i>	Area V of proboscis bare; dorsal cirrus short, attached to broad, quadrate lobe	Falcigers limited to notopodia	(Not known)	—

Name of Species	Unique characteristics	Diagnostic setae	Method of reproduction	Position of modification, if any
<i>Perinereis nigropunctata</i>	Area I with to 7 paragnaths; posterior notopodia prolonged in upper part	Falcigers limited to neuro-podia	Epitoky (Horst 1889 p. 171)	Male at 15/16; Female at 18/19
<i>Perinereis vancaurica</i>	Area VI of proboscis (see Chart IV)	Falcigers limited to neuro-podia	(Not known)	—
<i>Perinereis obtusata</i>	Area VI of proboscis with a single curved ridge	Falcigers limited to neuro-podia	Epitoky (Monro, 1931 p. 16)	Male at 13/14; Female at 17/18
<i>Perinereis helleri</i>	Area III of proboscis (see Chart IV)	Falcigers limited to neuro-podia	(Not known)	—
<i>Perinereis camiguina</i>	Area III of proboscis (see Chart IV); dorsal cirri short (Monro, 1931, p. 15)	Falcigers limited to neuro-podia	(Not known)	—
<i>Platynereis australis</i>	Natatory setae in female from setiger 30; homogomph falcigers present in some notopodia	Homogomph falcigers with long, slender appendage	Epitoky (Schmarda, 1861; Ehlers, 1904 and others)	Female at 30/31; Male at 19/20, or at 23/24 (see below)
<i>Platynereis dumerilii antipoda</i>	Acicula light to dark brown; dorsal and ventral cirri thick at base, slender distally; spinigers short-appendaged	Notopodia with homogomph falcigers and spinigers	Epitoky (see below)	Female at 22/23
<i>Platynereis magalhaensis</i>	Notopodial falcigers absent or inconspicuous	Notopodia with spinigers, neuropodia with spinigers and falcigers	Epitoky (Ehlers 1897)	Male at 20/21; Female at 25/26
<i>Platynereis polyscalma</i>	Transversely ribbed setae in far posterior segments; epitokes with greatly prolonged prostomium	Notopodial setae ribbed in distal ends	Epitoky (Chamberlin, 1919, p. 225)	Male at 14/15; Female at 22/23
<i>Platynereis bicanaledicata</i>	Notopodial falcigers conspicuous and usually dark at tip	Notopodia with simple falcigers; neuropodial falcigers composite	Epitoky (see below)	Male at 20/21 or 21/22
<i>Pseudonereis rotinestiana</i>	Posterior notopodial lobes long, rectilinear; areas V and VI of proboscis (see Chart IV) (Similar to <i>P. rotinestiana</i> )	Homogomph falcigers absent; heterogomph falcigers in neuropodia	(Not known)	—
<i>Pseudonereis anomala</i>		Homogomph falcigers present; heterogomph falcigers in neuropodia	Epitoky (Gravier, 1901, p. 195)	Female at 16/17

CHART IV.

PARAGNATHAL PROCESSES ON MAXILLARY (I-IV.) AND ORAL (V.-VIII.) RINGS OF THE PROBOSCIS

Name of species	Area I.	Area II.	Area III.	Area IV.	Area V.	Area VI	Areas VII & VIII
<i>Australonereis ehlersi</i> (See below)		Maxillary ring covered with long, slender papillae in one row					
<i>Ceratocephala edmondsi</i> (See below)	One long papilla	O	5 cirriform papillae in one row	O	O	One papilla	9 cirriform papillae in one row
<i>Ceratonereis mirabilis</i> (Augener, 1913, p. 170)	O	2 to 16 cones	About 10 cones	About 16 cones	O	O	O
<i>Ceratonereis labingensis</i> (Augener, 1913, p. 167)	O	7 to 10 in a double row	4 to 7 in a transverse rectangle	12 to 14 in a triangle	O	O	O
<i>Ceratonereis acquisetis</i> (Augener, 1913, p. 173)	3-4 in tandem, or 6-9 in a group	Many in a crescent	A broad group	A transverse group of 5-7 rows	O	O	O
<i>Ceratonereis erythraeensis</i> (Fauvel, 1919, p. 408)	2 to 6 small cones	2 curved rows with about 12 cones	Many in several irregular rows	Many in a group and continuous with II. and III.	O	O	O
<i>Ceratonereis peristomialis</i> (Knox, 1951, p. 223)	1 or 2 in tandem	3 rows in a transverse patch	3 rows in a transverse patch	A rectangular patch	O	Oval group of 2 or 3 curved rows	A continuous band of many paragnaths
<i>Micronereis halei</i> (See below)	O	O	O	O	O	O	O
<i>Namanereis quadrataiceps</i> (Benham, 1909, p. 242)	O	O	O	O	O	O	O
<i>Nicom aestuariensis</i> (Knox, 1951, p. 225)	O	O	O	O	O	O	O
<i>Neanthes vaalii</i> (Augener, 1913, p. 152, and below)	1 or 2 in tandem	10-12 in about 3 rows	About 25 in 2-4 rows	About 30 in a large group	3 in a triangle or 4	4 in a rectangle or 2-3 rows	A band of about 60 cones in 2-3 rows, continuous with VI.

Name of species	Area I	Area II	Area III	Area IV	Area V	Area VI	Areas VII & VIII
<i>Neanthes cricagnatha</i> (Knox, 1951, p. 217; Augener, 1913, p. 164)	About 4 in a square	About 30 in an oblique band	About 34 in a transverse cluster	About 40 in a tri- angular patch	V. to VIII. with a continuous band of many cones in 8 or 9 rows, or the rows number only 3-4 or 5-6.		
<i>Neanthes</i> near <i>cricagnatha</i> (See below)	About 9 in quadrate arrange- ment	About 14 in triangle	About 40 in a transverse cluster	About 30 in a tri- angular patch	V. to VIII. with a continuous band of many cones in many rows and a single row of larger cones on the maxillary side		
<i>Neanthes kerguelensis</i> (Augener, 1913, p. 165)	O	6-7 in an oblique double row	4-5 in a triangle	8-10 in a triangular patch	O	One only	3 in a transverse row
<i>Neanthes oryphoda</i> (Monro, 1938, p. 615)	2 in tandem	Oval patch of 3 rows	One only	Small patch of 2 rows	one cone	About 9 in a circular patch	A continuous sparse band of 3-4 rows
<i>Neanthes unifasciata</i> (Horst, 1924, p. 154)	one cone or none	14-18 in a double row	20-25 in 3-4 rows in a transverse group	About 25 cones in 6-7 rows	O	5-8 cones in a small transverse group	A row of 5 dis- tant cones
<i>Neanthes angusticollis</i> Augener, 1913, p. 149)	7-8 in an oval patch	At least 20 in an oblique patch	About 12 in a trans- verse oval group	About 25 in a large triangular group	O	6-9 in a rounded patch	A broad band, 5 deep midventrally to only 2-3 deep at the sides
<i>Eunereis marri</i> (Monro, 1939, p. 117)	O	O	O	O	V. and VI. with a con- tinuous band of about 4 rows	A continuous band of about 4 rows	A continuous band of about 4 rows
<i>Nereis fulcra</i> (Knox, 1951, p. 215) (not as in Willey, 1905) which gives:	O	2-3 curved rows	A variable cluster	A crescentic group	O	A small cluster of tiny cones	7-9 larger cones in a row or with variable number of tiny cones O
<i>Nereis denhamensis</i> (Augener, 1913, p. 158)	1 or 2 tandem	5 cones 13-16 in a double row	7 in 1 row 17-20 in a triple row	10 cones 17-20 or 24 in 3 or 4 rows	O	8 small cones in 2 obscure rows	About 8-10 or 13- 18 larger cones in a transverse row
<i>Nereis jacksoni</i> (Knox, 1951, p. 217; Kott, 1951, p. 95; Au- gener, 1913, p. 162)	O	2-3 curved rows, or a few, to only one	None or a trans- verse patch of 3-5-7 in a row, or triangle	A crescentic patch of 6-8-10 in 2-3 rows, or none	O	A small group or 1-4 in a row, or one, or none	A single row of about 7, or only 4 or 1-4 in a row, or none

Name of species	Area I	Area II	Area III	Area IV	Area V	Area VI	Areas VII & VIII
<i>Nereis thompsoni</i> (Kott, 1951, p. 104)	1 or 2 in tandem	2 oblique rows	4 cones with 3 in a row and 1 behind	About 20 cones in 2-3 irregular transverse rows	3 large cones in a triangle	4 cones in a rectangle	A row of larger cones and many smaller ones in about 6 rows behind
<i>Nereis peroniensis</i> (Kott, 1951, p. 101)	"Similar to that of <i>Nereis jacksoni</i> ."—Kott.						
<i>Nereis cockburnensis</i> (Augener, 1913, p. 155 and below)	O	8 in an oblique double row	2 in tandem	5-11 in a triangular patch of 2-3 rows	6 in 2 rows in an oval patch	5 large cones in a circle or cross	A broad band of many cones, 5-7 deep below and 2-4 deep at sides
<i>Nereis languida</i> (Kinberg, 1866, p. 169)	(Unknown)						
<i>Nereis robusta</i> (Quatrefages, 1865, p. 545)	(Obscure; the areas not clearly identifiable; paragnaths of maxillary ring much smaller than those of oral ring. Buccal ring with a broad band on ventral side and 2 lateral groups on dorsal side; maxillary ring with a band, and 3 groups above.)						
<i>Perinereis amblydonta</i> (Knox, 1951, p. 222)	2-4 cones in tandem	3-4 rows in a triangular patch	A triangular patch	A triangular patch	Usually 5 with a central one & 4 behind	A single curved ridge	About 50 cones in 2 irregular rows
<i>Perinereis variodentata</i> (Augener, 1913, p. 181 Kott, 1951, p. 112)	10 to many or only 3	About 9 in 2 oblique rows	Only 2, or a transverse patch of 2 rows	About 12 in 3 rows	6 cones in 2 rows	2 transverse ridges on each side	About 80 cones in 5-6 rows below, to only 1 row at the sides
<i>Perinereis vallata</i> (Knox, 1951, p. 219; Augener, 1913, p. 178)	1-3 cones	6-10 cones	About 14 cones in a cluster	About 20 cones in a cluster	1 set far back, or 3-4 in a triangle	7-9 or 8-20 in a transverse row	3 alternating rows of cones that may be flattened
<i>Perinereis brevicirris</i> (Knox, 1951, p. 220)	1-3 cones	A triangular cluster	Quadrangle patch with 1-3 cones on each side	A crescentic cluster	3 in a triangle or 4	10-20 conical or flat cones in a transverse row	3 irregular transverse rows or a few more
<i>Perinereis comissinoides</i> (Knox, 1951, p. 221)	1 large cone	Oblique group of 2-3 rows	3-6 smaller cones	Oblique group of 7-15 cones	3 large cones in triangle	2 (or 3) long transverse ridges	Up to 50 large and small cones in 2-3 rows

Name of species	Area I	Area II	Area III	Area IV	Area V	Area VI	Areas VII & VIII
<i>Perinereis barbara</i> (Monro, 1926, p. 317)	2 in tandem	Oblique patch of about 12 in 2 rows	Small group of 9 (or 14)	Large crescentic group	8 cones in a cross	A transverse ridge	A transverse band about 4 (to 8-9) deep
<i>Perinereis posuensis</i> (Augener, 1924, p. 351)	1 or 2 in tandem	10-11 in 2 rows, or only 5 cones	10 (to only 4) at middle and 1 isolated at each side	27 (or only 12-13) in 3-4 rows	one only	4 ridges on each side	25 cones in 2 rows widely separated from Area VI.
<i>Perinereis pseudocamiguina</i> (Augener, 1924, p. 340)	2 in tandem	8-10 in an oval patch	11 in 4 transverse rows in an oval group	27-28 in a triangular group	3 cones in a triangle	1 transverse ridge	About 42 cones in a band of 3 rows
<i>Perinereis calmanii</i> Monro, 1926, p. 319)	2 in tandem	An oblique double row	A small transverse group	About 3 rows in an oblique group	O	1 transverse ridge	An irregular row of 10 to 12 cones
<i>Perinereis nigropunctata</i> (Augener, 1922, p. 25)	7 in quadrate group	17 in a double or triple row	About 23 in an oval patch	About 23 in a crescent or in 4 rows	3 in a triangle	1 ridge distally pointed	About 33 in 2 rows, to only 1 row at the sides
<i>Perinereis vancouverica</i> (Augener, 1922b, p. 23)	2 in tandem	Many small cones	3 groups, a median large, and a pair or small, lateral ones	Many small cones	3 in a triangle	2 ridges on a side	2 broad bands, an anterior with larger and smaller cones, and a posterior with larger cones; continuous with VI.
<i>Perinereis obtusata</i> (Monro, 1931, p. 18)	5 cones in a cross	About 25 in a triple row	About 25 in a transverse patch	About 20 in an oblique patch	one large	1 ridge on each side	A double row
<i>Perinereis helleri</i> (Monro, 1931, p. 14)	2 in tandem	7 in subquadrate patch	3 groups; a median one of 9 cones, and 2 in tandem on each side	About 10 in an oval patch	2 in a triangle	A transverse ridge on each side	About 17 in an irregular double row
<i>Perinereis coniguina</i> (Monro, 1931, p. 15)			(As in <i>Perinereis helleri</i> , see above)				
<i>Platynereis australis</i> (Knox, 1951, p. 224)	O	O	4-5 interrupted lines of pectinae in a small transverse group	Several rows of pectinae (the most conspicuous area)	O	A small patch of 3-4 imperfect lines	5 small groups of minute pectinae



c	Name of species	Area I	Area II	Area III	Area IV	Area V	Area VI	Areas VII & VIII
	<i>Platymereis dumerilii</i> <i>antiboda</i> (See below)	O	O	As in <i>P. australis</i>	As in <i>P. australis</i>	O	2-3 close imperfect lines	as in <i>P. australis</i>
	<i>Platymereis magalhaensis</i> (Hartman, 1948, p. 60)	O	O	3 interrupted rows of pectinae	5 longer and 2 shorter rows of pectinae (the most conspicuous area)	O	3 long rows of fine pectinae	About 5 transverse, well-separated series of pectinae
	<i>Platymereis polyscalma</i> (Fauvel, 1932, p. 115)	O	O	pectinate clusters	pectinate clusters	none or one	A pectinate cluster	A double row
	<i>Platymereis bicanaliculata</i> (See below)	O	O	A broad area, 4-5 transverse rows of pectinae	7-8 rows of pectinae (see below)	O	2 irregular transverse rows of pectinae	A continuous narrow band of pectinae
	<i>Pseudonereis rotthestiana</i> (Augener, 1913, p. 186)	1 cone only	3-4 irregular rows of pectinae or part of another row	4 rows of many pectinae in an irregular rectangular area	4-5 rows of many pectinae or also 5 single cones	O	5-6 cones in a transverse row	About 16 to 19 large cones in a row, or in an alternating double row

(Similar to that of *Pseudonereis rotthestiana*, above.)

*Pseudonereis anomala*  
(Fauvel, 1922, 494;  
Kott, 1951, p. 93)

## KEY TO GENERA AND SPECIES

1	Parapodia uniramous	<i>Namanereis quadriceps</i>	
1	Parapodia biramous after the second parapodium		2
2	First segment with parapodia (fig. 18)	<i>Micronereis halei</i>	
2	First segment a smooth ring (fig. 12)		3
3	Peristomium prolonged forward ventrally to encompass the prostomium	<i>Cheilanereis peristomialis</i>	4
3	Peristomium not prolonged forward		4
4	Ventrum of anterior segments with rows of papillae (fig. 2)	<i>Australonereis ehlersi</i>	
4	Ventrum without rows of papillae		5
5	Proboscis with fleshy, cirrus-like papillae (fig. 13)	<i>Ceratocephala edmondsi</i>	
5	Proboscis with dark horny paragnaths on some or all areas		6
5	Proboscis without processes, its epithelium smooth or at most wrinkled	<i>Nicon aestuariensis</i>	
6	Paragnaths absent from oral ring of proboscis	<i>Ceratonereis</i>	11
6	Paragnaths absent from maxillary ring of proboscis	<i>Eunereis marri</i>	
6	Paragnaths typically present on both oral and maxillary rings of proboscis		7
7	Paragnaths in the form of pectinated rows on some or all areas		8
7	Paragnaths in the form of conical, separated processes		9
8	Maxillary ring with conical, and oral ring with pectinated processes	<i>Pseudonereis</i>	14
8	Both rings of proboscis with pectinated processes, or areas I, II and V usually bare	<i>Platynereis</i>	15
9	Area VI of proboscis with transverse ridges, or the ridges broken up into points in a straight transverse row	<i>Perinereis</i>	19
9	Area VI of proboscis with conical processes		10
10	Median and posterior notopodia with falcigers (fig. 30) or also spinigers	<i>Nereis</i>	20
10	Notopodia with spinigers; lacking falcigers	<i>Neonthes</i>	24
11	Prostomium deeply incised at midfront; dorsal cirri very long	<i>Ceratonereis mirabilis</i>	
11	Prostomium not incised in front; dorsal cirri not unusually long		12
12	Neuropodia with simple and composite falcigers	<i>Ceratonereis erythracensis</i>	
12	Neuropodia without simple falcigers		13
13	Dorsal and ventral cirri greatly reduced; area I of proboscis with 3-4 or to 6-9-pointed paragnaths	<i>Ceratonereis acquisetis</i>	
13	Dorsal and ventral cirri not greatly reduced; area I of proboscis bare	<i>Ceratonereis lapinigenis</i>	
14	Notopodia without homogomph falcigers	<i>Pseudonereis rothnestiana</i>	
14	Notopodia with homogomph falcigers in posterior segments	<i>Pseudonereis anomala</i>	
15	Notopodia with simple, heavy falcigers (fig. 38) in median and posterior segments	<i>Platynereis bicanaliculata</i>	
15	Notopodia without simple falcigers		16
16	Posterior notopodia with composite setae in which the appendage is strikingly ridged transversely	<i>Platynereis polyscalma</i>	
16	Posterior notopodia without setae that are distally ridged		17
17	Posterior notopodia with homogomph falcigers (fig. 37)		18
17	Posterior notopodia without falcigers or only an occasional inconspicuous one	<i>Platynereis mayalhaensis</i>	
18	Modified natatory parapodia in female present after segment 22	<i>Platynereis dumerilii antipoda</i>	
18	Modified natatory parapodia in female present after segment 30	<i>Platynereis australis</i>	
19	Area VI of proboscis with a single ridge on each side (Includes <i>Perinereis amblyodonta</i> , <i>barbara</i> , <i>calmani</i> , <i>helleri</i> , <i>camiguina</i> , <i>obfusca</i> , <i>nigropunctata</i> and <i>pseudocamiguina</i> . See Chart IV for distinguishing characteristics of each.)		
19	Area VI of proboscis with two ridges on each side (Includes <i>Perinereis camiguinoides</i> , <i>varioidentata</i> and <i>vancaurica</i> . See Chart IV for distinguishing characteristics.)		
19	Area VI of proboscis with four ridges on a side	<i>Perinereis ponuiensis</i>	
19	Area VI of proboscis with a continuous transverse series of cones that extends across areas V and VI (Includes <i>Perinereis vallata</i> and <i>brevicirris</i> . See Chart IV for distinguishing characteristics.)		
20	Notopodial lobe diminishes in size (fig. 26) in posterior segments		21
20	Notopodial lobe does not diminish in size posteriorly		22
21	Homogomph falcigers distally boldly bifid (fig. 27)	<i>Nereis jacksoni</i>	
21	Homogomph falcigers not boldly bifid (fig. 30)	<i>Nereis cockburnensis</i>	

22	Prostomium deeply incised at middle front	.....	<i>Nereis falcaria</i>	
22	Prostomium not incised at middle front	.....	.....	23
23	Notopodial falcigers boldly bifid	.....	<i>Nereis denhamensis</i>	
23	Notopodial falcigers not boldly bifid	.....	<i>Nereis peroniensis</i>	
24	Notopodial lobe comes to be very large on both sides of dorsal cirrus in medium and posterior segments	.....	<i>Neanthes oxypoda</i>	
24	Notopodial lobe does not come to be so large	.....	.....	25
25	Areas I and V of proboscis nearly or quite bare	.....	.....	26
25	Area I, or also V of proboscis with paragnaths	.....	.....	27
26	Notopodial lobe diminishes in posterior segments	.....	<i>Neanthes kerquelensis</i>	
26	Notopodial lobe does not diminish in size in back	.....	<i>Neanthes unifasciata</i>	
27	Area V of proboscis bare; area I with 7 or 8 cones in an oval patch	.....	<i>Neanthes angusticollis</i>	
27	Area V with paragnaths; area I otherwise	.....	.....	28
28	Area I was only 1 or 2 cones	.....	<i>Neanthes vaultii</i>	
28	Area I with many more cones	.....	.....	29
29	Areas V to VIII with a continuous band of many cones in 8 or 9 rows, to only 3 or 4 rows; area I with about 40 cones	.....	<i>Neanthes ericognatha</i>	
29	Areas V to VIII with a continuous band of cones, including a single row of larger cones on the maxillary side of the area; area I with about 9 cones	.....	<i>Neanthes</i> , near <i>ericognatha</i>	

### Australonereis, new genus

Type A. EHLERSI (Augener), 1913

This differs from other nereid genera most strikingly for having paired fleshy transverse ridges (fig. 2) on the ventrum of anterior segments. The armature of the proboscis consists of paired distal jaws and soft papillae on the maxillary ring only; the oral ring is bare. The first segment or peristomium is a smooth, apodous ring. The first two parapodial segments are uniramous; all others are biramous. Notopodia have spinigerous, composite setae and single acicula; neuropodia have spinigers and falcigers. In ovigerous adults the median and posterior segments have paired, papillar processes, presumably coelomostomes, located on the dorsal side of notopodia, within the base of the dorsal cirrus.

*Australonereis* approaches *Tylonereis* Fauvel (1911, p. 376) in its pharyngeal structures, but in the latter all setae are homogomph spinigers. *Leonnates* Kinberg (1866, p. 168) also has membranous processes on the proboscis, but they are limited to the oral ring, whereas the maxillary ring has horny paragnaths.

A single species, *A. ehlersi* (Augener) is referable to it.

### AUSTRALONEREIS EHLERSI (Augener) 1913

Fig. 1-11

*Nereis* (*Leonnates*) *ehlersi* Augener, 1913 pp. 142-145, pl. 3, fig. 53, text-fig. 12 a-c.

*Leonnates ehlersi* and *Leptonereis ehlersi* Monro, 1938, pp. 618-623, fig. 7-13.

*Locality*—Numerous individuals come from Lakes Entrance, Victoria, on the inner side of Ninety-mile Beach, where there is a considerable tidal current, and where the water is marine, from a sand spit uncovered at low tide; the worms form beds and occupy U-shaped tubes in which the ends are uncovered (observations by Miss Barbara Dew).

*Australonereis ehlersi* has remained known only through sparse catches from the Swan River area, Western Australia. The present numerous individuals come from Lakes Entrance, Victoria. They are conspicuously large, measure to 140 mm. long and 12 mm. wide with parapodia. The body is greatly depressed, especially in its median and posterior regions. The following description is based on specimens from Victoria.

On the everted proboscis the oral ring is dusky and smooth or irregularly rugose; it lacks processes. The maxillary ring is pale and has a continuous band of many, more than 50, short, cirrus-like processes in 3 to 5 irregular rows.

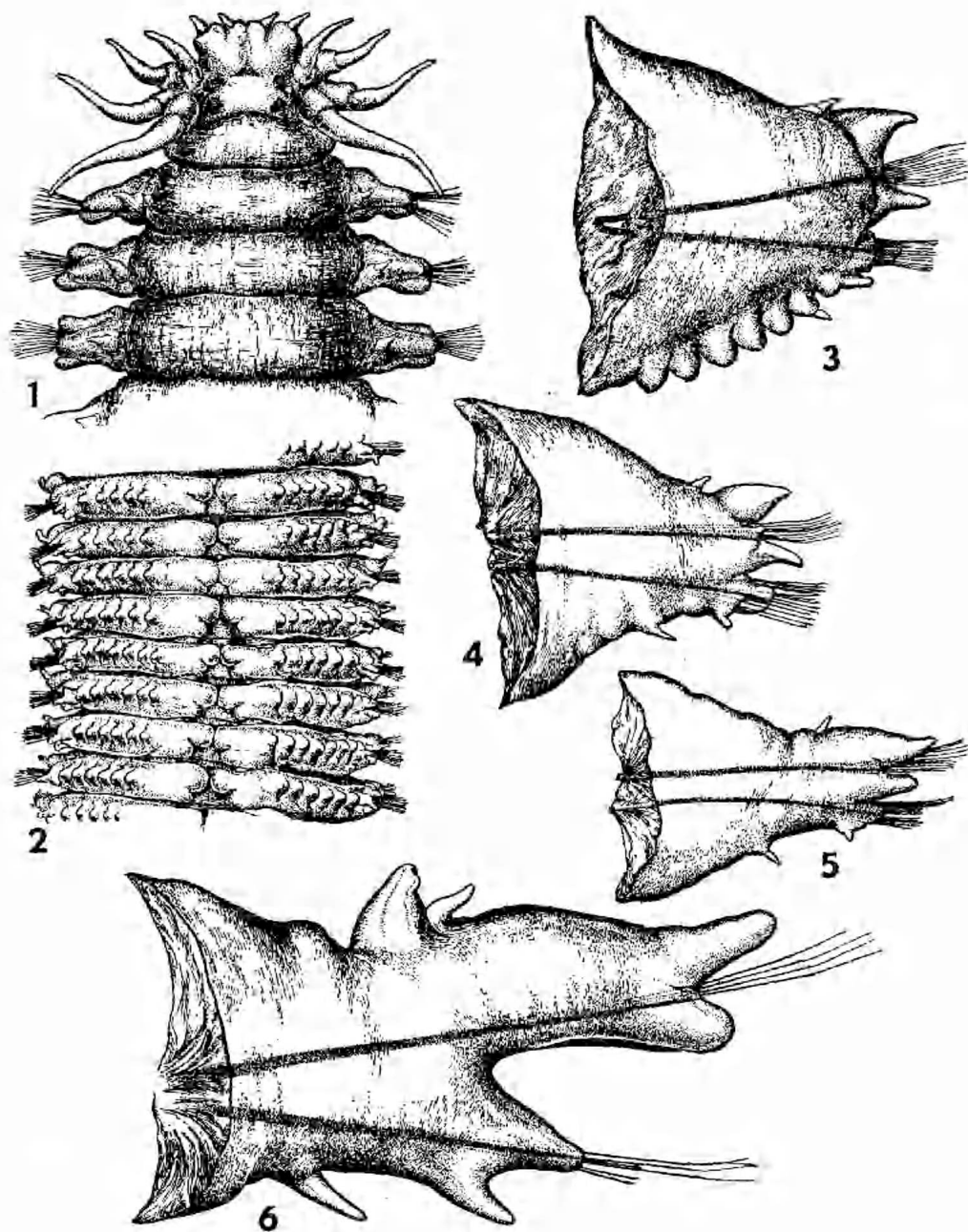


Fig. 1-6. *Australoneris ehlersi*

1. Anterior end in dorsal view, including first 3 setigers,  $\times 10.5$ .
2. Ventral side of body showing segments 10 to 17,  $\times 5.2$ .
3. Anterior parapodium from papillated region, seen from the front,  $\times 10.5$
4. A parapodium from the middle region of the body, seen from the front,  $\times 10.5$ .
5. A slightly more posterior parapodium seen in posterior view,  $\times 10.5$ .
6. A far posterior parapodium showing the large coelomostome adjacent to the small dorsal cirrus,  $\times 51$ .

They are fewest at the sides and most numerous midventrally; those on the oral end are slightly larger than those on the maxillary one. They are limited to a band that separates maxillary and oral parts, giving the impression, especially on the retracted proboscis, of being oral, not maxillary (hence Augener's observation that they are oral). The jaws are translucent, light yellow to horny brown distally; they have 7 to 9 short, oblique teeth along the cutting edge.

The first 6 parapodial segments have each a slender ventral cirrus on a papillar elevation. From the seventh segment the ventrum has an additional elevation within the base of the ventral cirrus and on the next 9 or 10 segments these papillations increase to about 6 or 7 on a side (fig. 2). The ventrum in this region is rugose. After segment 30 the papillations diminish rapidly and are absent from posterior segments.

Parapodia of the first 17 to 19 segments differ from those farther back in that their distal lobes (both notopodia and neuropodia) are thick and glandular. The glands are most conspicuous at anterior sides of parapodia and reach their maximum thickness and extent in segments 10 to 18, where the uppermost lobe comes to be transversely rugose and resembles the furrows of the ventrum in the same region. After about segment 20 these parapodial areas are abruptly absent. Dorsal and ventral cirri are slender, short and inconspicuous; they are simple and tapering throughout the body.

Setae are in thick, yellow fascicles and most numerous in anterior segments. Those in notopodia are entirely spinigerous (fig. 11). Neuropodia have both spinigers and falcigers (fig. 7-10). The latter have a cutting edge with a single series of denticles (fig. 7, 9); they terminate in a curved process that is bounded by a series of denticulations continuous from the cutting edge. Acicula occur singly in parapodial rami; each is a slender, distally tapering, straight black rod; the deeply embedded base is pale.

In postmedian segments, from about segment 50 in shorter, to about segment 68-70 in longer, individuals, there is present, immediately within the base of the dorsal cirrus, a papillar organ which comes to increase in size to surpass that of its corresponding dorsal cirrus (fig. 6); its distal end is penetrated by a pore. By means of microtome sections<sup>(1)</sup> it is possible to trace ducts which penetrate these papillae, and to follow their course into the coelomic spaces. Occasionally one can find larger ova in the cut. It can hardly be doubted but that these are coelomoducts which function at maturity for release of gonadal products. Whether primitively retained from ancestral stages, or secondarily derived might be determined from a study of the development of this species. Among the numerous individuals examined, I have found only ovigerous ones, all showing the coelomostomes present from an anteromedian region to the posterior end of the body.

In this connection it is interesting to recall a statement by the late E. S. Goodrich (1945, p. 173): "In species of Nereidae, co-existing with metanephridia are a pair of specialized coelomostomes, the so-called 'dorsal ciliated organs.' . . . They occur in all species . . . but may vary somewhat in size. They appear in the young, persist throughout life, though in the heteronereid phase they are usually reduced or absent. That this 'dorsal ciliated organ' is indeed the representative in the Nereidae of the coelomoduct or genital funnel of the Capitellidae and other Polychaeta, . . . there can now be no doubt . . . But it has lost its original genital function in the Nereidae, no longer requires an opening to the exterior, and has become converted into a 'ciliophagocytal organ,' at all events in the majority of species in which the genital products are known to escape by dehiscence . . . It is possible, however, that some species still exist which have no specialized epitokous stage, and that in them the coelomostomes still function as genital ducts."

<sup>(1)</sup> I am indebted to Mr. Donald J. Reish for the preparation of the sections.

It seems probable that *Australonereis ehlersi* is indeed such a species in which the dorsal ciliated organ is replaced by the coelomostome, and that it functions as a genital duct, acquiring an opening to the exterior. There is no indication of epitoky or parapodial transformation in the individuals that have been examined.

The pygidium is a terminal, dark brown collar; a pair of long, cirriform processes is inserted ventrally; each is about as long as the last 10 segments.

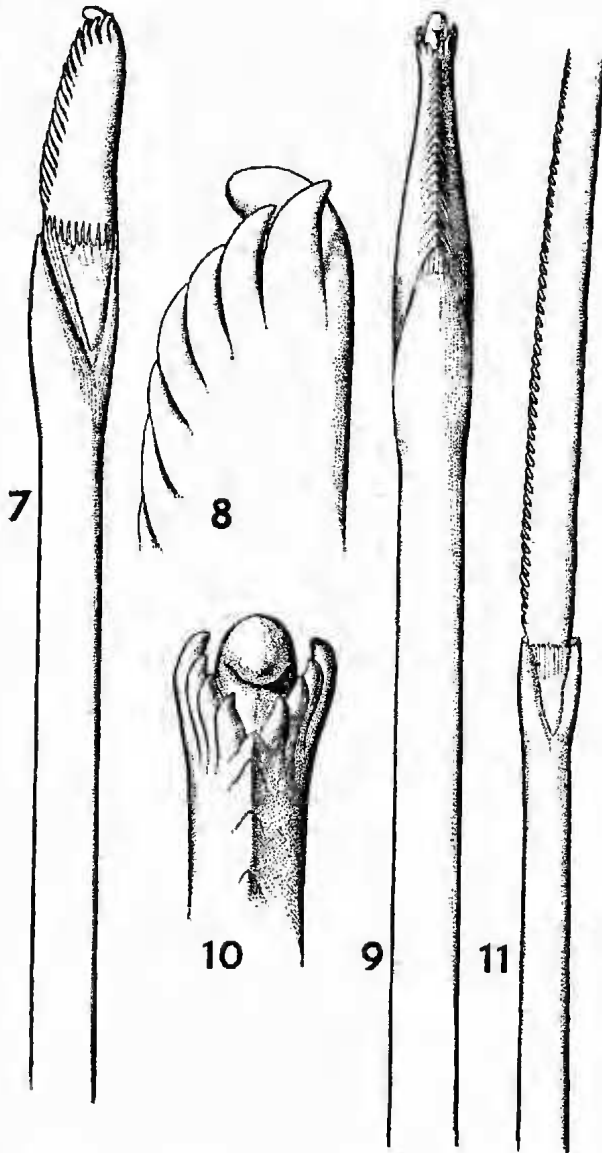


Fig. 7-11

*Australonereis ehlersi*

7. Neuropodial falciger seen from the side, x 521.
8. Distal end of a neuropodial falciger showing details of cutting edge and terminal fang, x 2010.
9. Neuropodial falciger seen from the cutting edge, showing arrangement of single row of denticulations, x 521.
10. Distal end of a neuropodial falciger seen from the cutting edge, x 2010.
11. Portion of a homogomph spiniger seen from the side, the tapering pointed tip not shown, x 521.

*Australonereis ehlersi* was first assigned to the genus *Leonnates* (Augener, 1913) and later to *Leptonereis* (Monro, 1938, p. 618). Augener thought that the oral ring is papillate; Monro found the maxillary ring to have papillae. Augener found no falcigers above the neuroaciculum; Monro found them in anterior segments; Augener called the falcigers heterogomph; Monro said they are nearly to quite homogomph. These discrepancies can readily be attributed to subjective interpretations. The species cannot be assigned to *Leonnates* Kinberg or

*Leptonereis* Kinberg. In the first the maxillary ring of the proboscis has horny paragnaths; in the second the proboscis rings are both bare.

*Australonereis ehlersi* is now known from opposite sides of the southern half of Australia, at Swan River, Western Australia and Lakes Entrance, Victoria.

CERATOCEPHALA Malmgren, 1867, emended

Type C. LOVENI Malmgren

The generic diagnosis is here expanded to include species in which the pharyngeal papillar processes are present on both rings of the proboscis instead of only the oral ring (see Hartman, 1952, pp. 15-18, for detailed account).

***Ceratocephala edmondsi*, n. sp.**

Fig. 12-17

*Locality*—American River, Kangaroo Island, South Australia; very common in the sand of a cockle (*Katclysia* sp.) bank (9 specimens), coll. S. J. Edmonds.

Length of a larger, posteriorly incomplete, individual is 27-39 mm.; width at the widest (anterior) part is 3-4 mm.; number of segments is more than 60. The general colour (preserved) is pale with melanistic spots on dorsal and ventral sides; it resembles that of species of *Platynereis*. The prostomium has two pairs of eyes that are large, subequal, in trapezoidal arrangement; the anterior ones are wider apart. The proboscis (everted) shows the following parts: area I (fig. 12) has one papilla, II and V are bare; III and IV together have 5 cirriform papillae in a transverse row; VI has a single papilla on a side; VII and VIII have 9 cirriform papillae in a transverse row (fig. 13). Jaws are thin, translucent, horny brown; they have 7 to 9 shallow crenulations at the cutting edge.

The first 2 parapodia on a side are uniramous; each has composite spinigers and falcigers; succeeding parapodia are biramous. From the third a notopodium is developed and has a full fascicle of composite spinigers. At the eighth or ninth notopodium there are 15 to 20 spinigers and single black acicula. Neuropodia have a supra-acicular bundle of about 10 spinigers and 9 falcigers, and single black acicula that taper distally and are turned upward at the tip. The sub-acicular setal bundle has about 14 falcigers (fig. 16) and 7 spinigers (fig. 17). Dorsal and ventral cirri are simple throughout. A fiftieth parapodium is shown in fig. 14 and an eighth one in fig. 15.

The habitat is sandy beaches in which cockle shells occur; the nereid occupies a sandy tube constructed with a thin, gelatinous matrix (observation by Mr. S. J. Edmonds).

*Ceratocephala edmondsi* differs from other species of the genus in that the maxillary ring of the pharynx has papillae instead of lacking them; ventral cirri are simple throughout, instead of double on some or all segments. The genus is a small one, known for only 5 or 6 other species or subspecies (Hartman, 1952, p. 19) from widely scattered parts of the world. *C. edmondsi* is the only one known from Australia. *C. sibogae* Horst, off Dutch East Indies, is the nearest in geographic range. It is clearly separable from *C. edmondsi* in its pharyngeal processes in that the former has papillae nearly absent, with only 2 present on area V.

It is a pleasure to dedicate the species to its collector, Mr. S. J. Edmonds of the University of Adelaide, South Australia.

*C. edmondsi* is known from only one locality, American River, Kangaroo Island, South Australia, littoral.

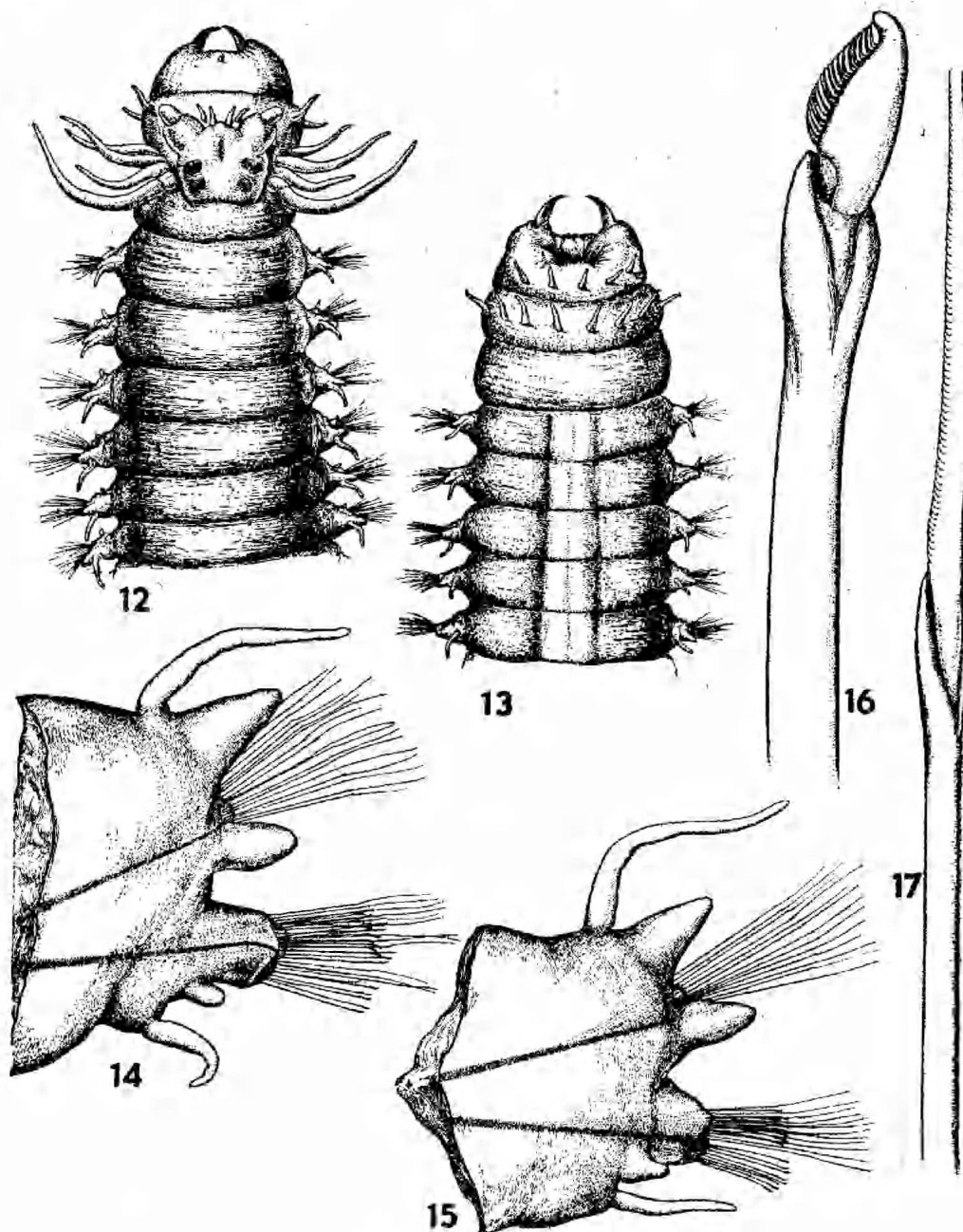


Fig. 12-17. *Ceratocephala edmondsi*

12. Anterior end with everted proboscis and first 6 setigers, in dorsal view, x 15.  
 13. Anterior end with proboscis everted, in ventral view, x 15.  
 14. Fiftieth parapodium seen from the front, x 45.  
 15. Eighth parapodium seen from the front, x 45.  
 16. Neuropodial falciger from a posterior parapodium, x 700.  
 17. Articulating portion of a spiniger, seen from the side, x 700.



## MICRONEREIS Claparède, 1863

Type *M. VARIEGATA* Claparède*Micronereis halei*, n. sp.

Fig. 18-21

*Locality*—Sellick Beach, South Australia, at outer edge of reef, 16 January 1936, at low tide from stones in rock pools (12 individuals), coll. Mr. H. M. Hale and Mr. K. Sheard.

This is a small, white species, greatest length is about 7 mm.; width 0.55 mm. without and 0.85 mm. with parapodia. Number of segments is 20 to 25. The prostomium is broadly quadrate (fig. 18); its posterior margin is clearly marked off from the first segment. There are 2 pairs of lenticulated eyes, with the anterior pair slightly larger and wider apart than the posterior one; all are similar in that the basal part is dark red and there is a large, spherical pale lens. The frontal margin of the prostomium is weakly indented, and the small oval paired palpi can be seen only by viewing the prostomium from below. There are no prostomial frontal antennae. The four pairs of tentacular cirri are directed forward and outward; all are similar with slight variation in length; the antero-ventral pair are shortest and the dorsal posterior pair are longest. All 8 are on short bases (not shown in fig. 18). Each has a slight subdistal swelling, diffusely brown in colour, with a simulated articulation just below the brown pigment.

The first segment has uniramous parapodia in which the setal lobe is long, compressed, directed laterally; it has a cirriform ventral cirrus that is attached near the middle of the parapodial base; it extends distally not as far as the lobe. Stumps of 8 to 10 slender setae and single acicula are visible. The second parapodium is similar to the first but a little larger.

The third and successive segments have biramous parapodia. Notopodia and neuropodia are widely separated from each other. The dorsal cirrus is a long, cirriform process at the upper, outer edge; the ventral cirrus is similar but somewhat shorter and attached near the middle of the lower base of the parapodium. In addition, both notopodia and neuropodia have a long, digitate lobe that extends distally, attached one at the inferior outer edge of the notopodium, the other at the superior outer edge of the neuropodium (fig. 20). These lobes resemble dorsal and ventral cirri but they are not so thick and extend laterally not quite as far as the cirri. Acicular lobes are compressed, broadly triangular and have an acute tip.

All setae are homogomph spinigers (fig. 2) with the longest appendages several times as long as the shortest ones. The uppermost have the longest appendage and the length diminishes gradually ventrally. Notopodia have 15 to 25 spinigers and single yellow acicula. Neuropodia have 15 to 20 spinigers and single yellow acicula.

The pharyngeal apparatus (seen only by dissection since the proboscis was not everted on any individual) is a subspherical, muscularized mass. It connects distally with the mouth and proximally with the thin-walled, alimentary tract. There are no paragnaths. A pair of large translucent, yellow jaws are inserted, one on either side of the muscular tissue. Each jaw is broadly oval at the base and continued distally to end in about 6 triangular teeth along the concave cutting edge (fig. 19). If the jaws are dimorphic in this species as they have been described for *M. variegata* Claparède (Racovitza, 1893), it may be presumed that the description is based on the jaws of a female individual.

The only other known species of the genus is *Micronereis variegata* Claparède, from the Mediterranean Sea, more widely recorded from western Canada (Berkeley and Berkeley, 1948, p. 60), though with some doubt. *M. variegata* Claparède differs from *M. halei* in that digitate lobes are lacking from the inner,

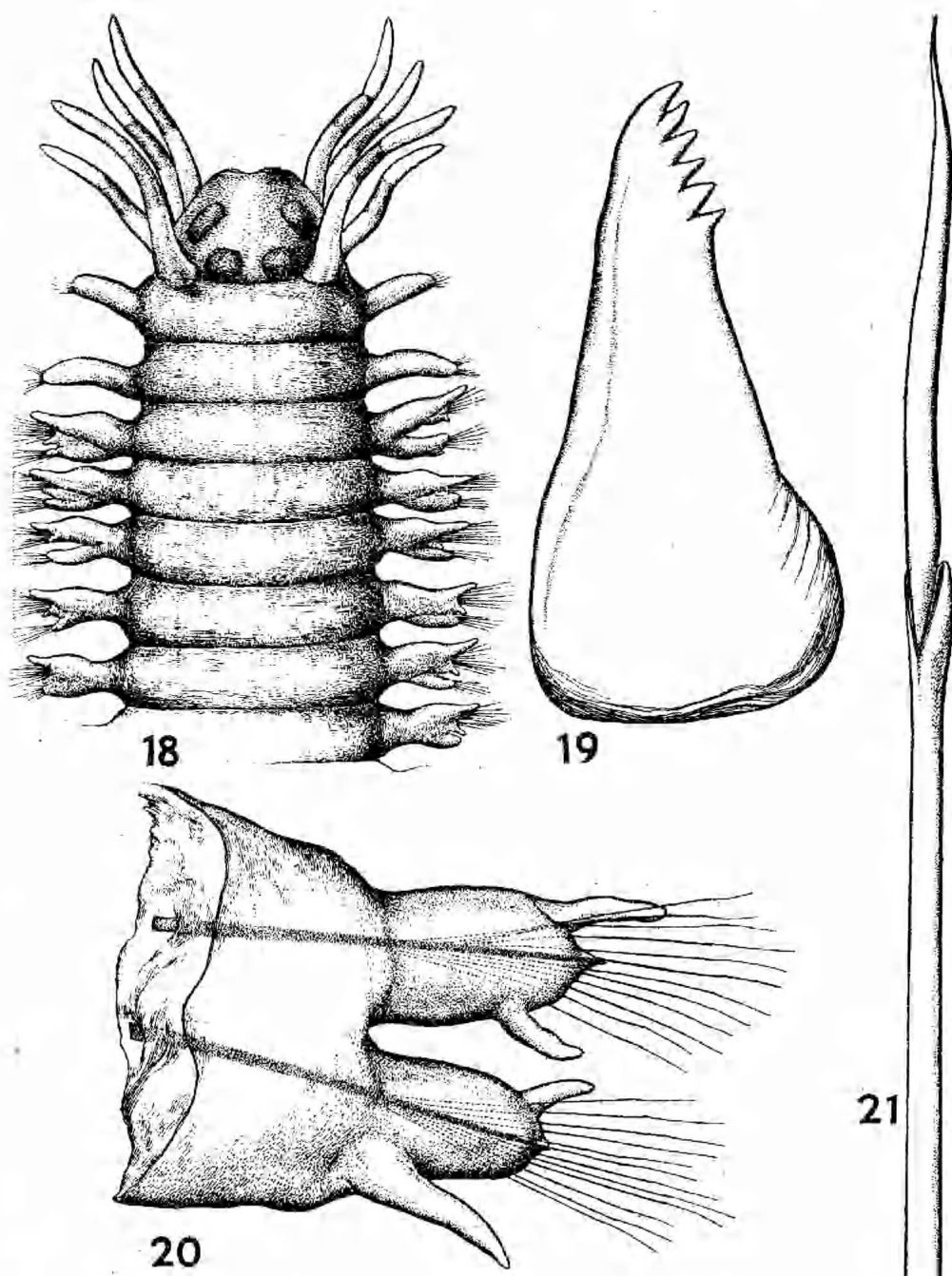


Fig. 18-21. *Micronereis halei*

18. Anterior end in dorsal view, proboscis retracted, x 94.  
 19. An entire jaw plate showing distal toothed edge and embedded part, x 417.  
 20. A median parapodium in posterior view, x 62.5.  
 21. Spinigerous seta from a median parapodium, x 1638.

proximal margins in the first; the pharyngeal jaws lack the broad base, and the first parapodia have conspicuous tufts of setae. Prostomial and peristomial structures also differ (see Fauvel 1923, pp. 332-333, for illustrated account).

It is a pleasure to name this species for its collector, Mr. Herbert M. Hale, Director of the South Australian Museum.

*Micronereis halei* has been found only from littoral zones in South Australia.

NAMANEREIS Chamberlin, 1919

Type N. QUADRATICEPS (Blanchard)

NAMANEREIS QUADRATICEPS (Blanchard), 1849

*Lycastis quadraticeps* Benham, 1909, pp. 242-244, pl. ix, fig. 2-10.

This brackish nereid was first described from Chile. Benham (1909, p. 244) recorded it from Campbell Island, on shore near the exit of a creek from the flank of Mount Honey; the shore above high-water mark is traversed by numerous little watercourses oozing through the earth above (Benham); also in sea pools. Benham's detailed description compares so favourably with individuals I have examined from southern and central California, from a similar brackish niche, that specific identity seems probable. The only differences I can find are these: The pygidium shown by Benham (his fig. 5) as a constricted collar with a pair of divergent lateral processes, is shorter and has a longer ventral lobe with the anal aperture between the upper and lower parts; the ventral portion has a pair of small, oval papillae inserted at the distalmost margin. Neuropodial falcigers are shown by Benham with a single series of subequal crenulations at the cutting edge; I see a single row of teeth that are longest near the base and diminish in size to near the distal third. The pharyngeal jaws have teeth that are long, sharp, obliquely inserted. These differences may have no specific importance.

*Namanereis quadraticeps* may be expected to occur in high intertidal zones of the southern shores of Australia; it should be sought especially in zones where there is only a light spray of sea-water.

NEANTHES Kinberg, 1866

Type N. VAALII Kinberg

The collections have made possible an examination of the type species from the type locality, for a genus which is widely represented in littoral zones of the Northern Hemisphere. Both atokal and epitokal individuals of both sexes are represented. The account below is based on these collections.

NEANTHES VAALII Kinberg, 1866

Fig. 22-25

*Neanthes vaalii* Kinberg, 1866, p. 171.

*Nereis albanyensis* Augener, 1913, pp. 149-153, pl. ii, fig. 6, text fig. 14.

*Neanthes vaalii* Augener, 1922, pp. 20-21.

*Localities*—American River, Kangaroo Island, in mud flats (5 individuals), coll. S. J. Edmonds; Port Adelaide, in tidal river (2 male and 8 female epitokes), coll. S. J. Edmonds; Rushcutters Bay, Port Jackson, scraped off hull of a yacht, 6 Oct. 1950, (1), coll. B. Dew; Athol Bight, public jetty, off piles, 12 Oct. 1950, (2), coll. B. Dew; Milsons Point, Port Jackson, off piles and mooring chains, 23 Oct., 1950, (3), coll. B. Dew; Venus Bay Inlet, Eyre Peninsula, South Australia, associated with clusters of *Modiolus*, (3), coll. S. J. Edmonds; Point Wynyard, north-west Tasmania, Apr. 1936 (4 tiny individuals), coll. H. M. Hale and N. B. Tindale.

Preserved, the pigment pattern resembles that of *Platynereis* species in having dark segmental spots over the dorsum and parapodia. Length of atoke individuals (preserved) is 70 mm. Notopodia have spinigers only; neupodia have a supra-acicular fascicle of homogomph spinigers and heterogomph falcigers, and a sub-acicular fascicle of heterogomph spinigers and heterogomph falcigers (fig. 25). There are no notopodial falcigers.

In male epitokes the first 7 segments have thickened dorsal cirri; the first 18 segments are otherwise unmodified, or the eighteenth is slightly changed with a few accessory lobes. Natatory setae are present from segment 19. Natatory parapodia are present to the end of the body; the pygidium has a rosette of many similar, slender papillae. Overall size is somewhat less than that for the female which measures to 50 mm. long.

Typical natatory parapodia (fig. 24) have dorsal cirri that are crenulate, and accessory lobes. In epitokal females modified natatory setae are present from segments 21 to 89; parapodia from 90 to 102 (posterior end) differ in having only single dark acicula in each ramus (setae lacking); the body terminates in a pygidium with a constricted smooth collar.

In mature individuals the 4 prostomial eyes are enlarged, arranged in a rectangle; each is a circular convex disk, purplish red at the periphery, fading centrally; each has a tiny white circular lens.

In atokal individuals the notopodia lack conspicuous preacicular and post-acicular lobes such as characterize northern representatives of the genus *Neanthes*, notably *N. virens* (Sars) and *N. brandti* (Malmgren). Median (fig. 22) and posterior (fig. 23) parapodia are similar to one another.

The pharyngeal processes (based on a female epitokal individual from Port Adelaide) are arranged thus: I has 2 cones in tandem; II has 10 cones in a triangular area; III has about 22 cones in a broadly oval patch; IV has a large crescent of about 30 cones of larger and smaller cones; area V has 3 cones in a triangle; VI has 3 cones in a transverse line; VII and VIII (continuous) have 2 or 3 irregular rows of 30 or more cones.

*Nereis albanensis* Augener, 1913, p. 152, from Western Australia has been referred to *Neanthes vaalii* Kinberg (Augener, 1922, p. 20). *Nereis* (*Neanthes*) *albanensis* Kott (1951, p. 106) from Point Peron, Western Australia, is another species and belongs to the genus *Nereis*, s. s. since there are dorsal falcigers in notopodia.

The distribution of *Neanthes vaalii* is indicated in Chart II.

#### NEANTHES, near CRICOGNATHA (Ehlers) 1905

*Nereis cricognatha* Ehlers, 1905, p. 29; Augener, 1913, pp. 163-164.

*Neanthes cricognatha* Knox, 1951, pp. 217-218, pl. 45, fig. 6-8; Fauvel, 1947, p. 8.

*Nereis arenaceodentata* Benham, 1916, p. 134, pl. 46, fig. 1-3.

*Localities*—American River, Kangaroo Island, (2), coll. S. J. Edmonds; Port Adelaide, outer harbour pilings, sublittoral fouling materials, (1), coll. S. J. Edmonds; Sellick Beach, South Australia, on edge of reef permanently covered, (2), coll. H. M. Hale.

Length attains about 30 mm. Notopodia lack homogomph falcigers, thus this is regarded as a species of *Neanthes*. Parapodial lobes are bordered with a dark, glandular margin. On the proboscis both oral and maxillary rings have complete circlets of many paragnaths.

The present individuals differ from *Neanthes cricognatha* previously recorded (see synonymy above) in that areas V to VIII of the proboscis have a circlet of larger cones on the maxillary side, and 4 to 7 rows of uniformly much smaller cones on the oral side.

The distribution of the stem species is indicated in Chart II.

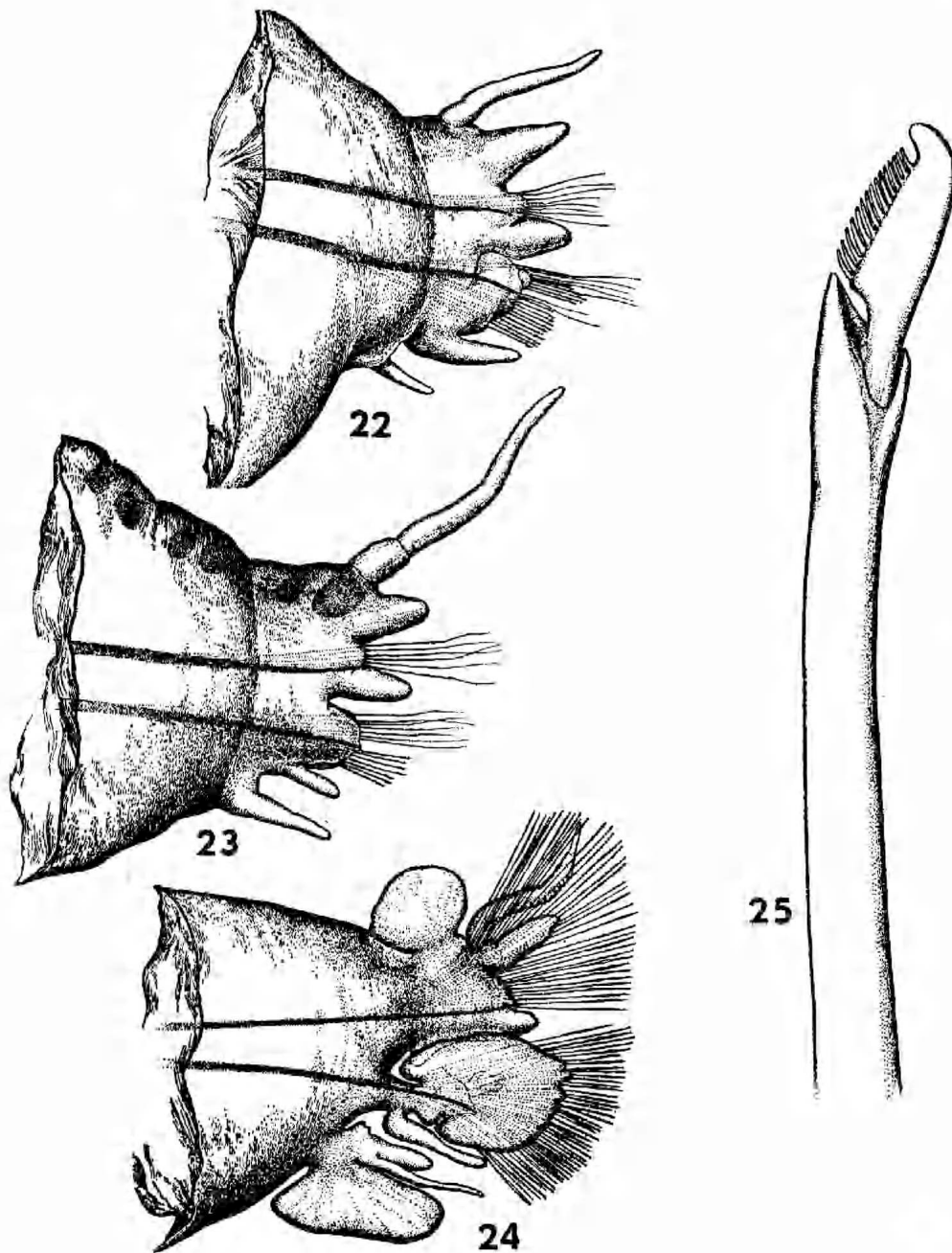


Fig. 22-25 *Neanthes vaalii*

22. A median parapodium in anterior view, showing maximum development of acicular lobe,  $\times 35.7$ .  
 23. Fifteenth last parapodium in anterior view,  $\times 63.2$ .  
 24. Thirtieth parapodium from an epitokous male specimen,  $\times 15.7$ .  
 25. Falcigerous neuropodial hook from an unmodified parapodium,  $\times 658$ .

## NEANTHES KERQUELENSIS (McIntosh), 1885

*Nereis kerguelensis* McIntosh, 1885, pp. 225-227, pl. 35, fig. 10-12, pl. 16a, fig. 17, 18; Fauvel, 1916, p. 433; Benham, 1916, p. 122.

*Localities*—Port Willunga, S. Aust., 18 Nov., 1945, (2), coll. S. J. Edmonds; and Sellick Beach, S. Aust., on edge of reef, Jan. 1936, (1), coll. H. M. Hale.

Notopodia have spinigers only, thus this is referred to *Neanthes* Kinberg. On the proboscis areas I and V are bare; VI has one cone on a side; VII and VIII have a single row of 9 cones; each of areas II, III and IV has a heap of small paragnaths. In posterior parapodia the notopodial lobe diminishes in size and is far surpassed by the acicular lobe of the same segment.

*Nereis kerguelensis oligodonta* Augener (1913, pp. 164-166), from Western Australia, also lacks homogomph falcigers and is presumably a species of *Neanthes*. It differs from the stem species in having only 3 cones in a transverse row on areas VII and VIII.

## NEANTHES ANGUSTICOLLIS (Augener), 1913

*Nereis angusticollis* Augener, 1913, pp. 145-149, pl. 2, fig. 14, text fig. 13.

*Not* Kinberg, 1866, p. 169.

Notopodial falcigers are absent, thus this is referred to *Neanthes* Kinberg. On the proboscis area I has 7 or 8 cones in an oval heap, II has at least 20 in an oblique triangular area; III has about 12 in an oblique oval group; IV has about 25 cones in a triangle; V is bare; VI has 9 or 6 in a rounded group; VII and VIII form a broad transverse band with 2 or 3 to 5 rows, the band widest midventrally and narrowing toward the ends. Acicula are black and occur singly in parapodial bases.

*Nereis angusticollis* Kinberg (1866, p. 160) from Tahiti is a *Nereis*, sensu stricto, since there are homogomph falcigers in notopodia.

## NEREIS LINNAEUS, 1758

Type *N. PELAGICA* Linnaeus

The species of *Nereis* from the southern half of Australia are peculiar for having several in which the notopodial lobe diminishes in size going back, and homogomph falcigers have a large lateral tooth near the apex. These characters are known for *N. falcaria*, *N. jacksoni*, *N. denhamensis* and *N. thompsoni* (see below). In others the oral ring is nearly to quite bare, approaching a condition in species of *Ceratonereis* Kinberg; such are the species *N. jacksoni* and *N. falcaria*. Others have tentacular (buccal) cirri that are annulate, as in *N. cockburnensis*. These features are neither generic nor limited to Australian species, but are more frequently encountered among species from the southern hemisphere than elsewhere. Thus, the coarsely toothed homogomph falciger is known for *N. sonata-persica* Fauvel from Persia, and for *N. funchalensis* Langerhans from Madeira. The posterior notopodial lobe diminishes in size in *Neanthes kerguelensis* (McIntosh) (see Ehlers, 1897). Annulation of tentacular cirri is encountered in other species and genera, notably *Nereis eugeniae* (see Ehlers, 1897), *Nereis angusta* Kinberg (1866), *Neanthes kerguelensis* (see Ehlers, 1897), *Neanthes ruficeps* (Ehlers, 1905) and *Platynereis australis* (see Ehlers, 1905), all from the Southern Hemisphere.

The several species discussed below are those which have occurred in greatest abundance and for which some details have been obscure.

## NEREIS DENHAMENSIS Augener, 1913

*Nereis denhamensis* Augener, 1913, pp. 156-159, pl. 3, fig. 51, text fig. 16; Fauvel, 1922, p. 494; Kott, 1951, pp. 99-101, fig. 3, 4.

Homogomph falcigers first appear after segment 20 to 30 and number 3 or 4 in a fascicle; they are thicker than their accompanying spinigers. The falcate appendage is short, weakly curved and projects from the end of the shaft for only about half its length; the cutting edge has 2 or 3 small teeth.

On the proboscis area I has 1 or 2 cones in tandem; II has about 12 cones in 2 rows; III has about 12 cones in 3 rows; IV has 12 to 15 cones in a triangular patch; V has none; VI has 8 to 10 in an oval patch of 2 or 3 rows; VII and VIII have a single row of 8 to 10 larger cones.

In epitokal male individuals the parapodial change to natatory condition is at segment 15. Dorsal cirri of modified segments are sharply geniculate in their distal extremity and crenulate along the outer margin of the basal part.

One character named by Augener (1913, p. 158) but not commented on further, states that: "An den vorderen Rudern mit dorsalen Grätenborsten ist die Spitze der ventralen Sicheln gedeckt." If this indicates the presence of a hooded condition of anterior neurofalcigers, it describes a character unique for this species.

See Chart II for distribution.

#### NEREIS JACKSONI Kimberg, 1866

Fig. 26-29

*Nereis jacksoni* Kimberg, 1866, p. 169; Augener, 1922, pp. 27-30, fig. 6; Augener, 1927, pp. 130-133; Knox, 1951, pp. 216-217; Kott, 1951, pp. 95-98; fig. 3. *Nereis heurissonensis* Augener, 1913, pp. 159-163, pl. 3, fig. 52, text fig. 17.

*Localities*—Sellick Beach, Sth. Aust., on edge of reef permanently covered and at low tide, Jan. 1936, (1), coll. H. M. Hale and K. Sheard; Shell Point, Botany Bay, N.S.W., from a 6-month fouling plate, estuarine, Feb. 1947, (4), coll. B. Dew; Cape Cove, Port Jackson, N.S.W., dredged in 3-4 fms. from a gritty bottom, Oct. 1950 (4), coll. B. Dew; Hungry Point, Cronulla, N.S.W., under rocks, Sept. 1950, (2), coll. B. Dew.

The proboscis has few paragnaths; areas I and V have none; II and III have a few cones and IV has a few more; VI has 1 to 4 only; VII and VIII have a single row of only 2 to about 7 cones (see also Chart IV). Jaws are dark amber in colour, thin, and have 5 or 6 oblique teeth at the cutting edge.

Prostomial antennae are long; they extend forward to near the distal end of the palpi. Peristomial cirri are short, the longest reaches back only to about the second setigerous segment and others are shorter; all are irregularly annulated. The 4 eyes are embedded and visible through the smooth epithelium; the 2 of a side are nearer together but widely separated from those of the opposite side; the anterior ones are the larger. Each eye has a reddish purple iris and a large pale to white lens, nearly or over half as large as the diameter of the eye. The anterior margin of the prostomium is entire, not incised.

In posterior segments the notopodial, or supra-acicular, lobe diminishes (fig. 26) conspicuously in size but is visible as a distinct lobe to the end of the body. Homogomph falcigers are present in median and posterior segments; their earliest presence varies from the fourteenth, or not before segment 17 or 18. They have an appendage (fig. 27, 28) that is short, distally bifid; those in front are similar to those behind or the latter may lack the basal-most teeth (fig. 29).

Some individuals from Shell Point, Botany Bay, taken 2 Feb. 1941, are ovigerous, with large ova crowding the body cavity from the third setiger more posteriorly. There are no signs of epitoky, such as the presence of modified lobes or specialized setae. Indications are that development is direct. This is in contradiction to what Kott (1951, p. 97) found for individuals from Western Australia. Augener (1913, pp. 159-60) examined about 50 specimens taken from May to September and found them all atokal.

The more extended distribution is indicated in Chart IV.

## NEREIS THOMPSONI Kott, 1951

*Nereis (Neanthes) thompsoni* Kott, 1951, pp. 103-105, fig. 5.

This is here referred to *Nereis* since notopodia have homogomph falcigers. It bears resemblance to *Nereis denhamensis* (see above) but differs in its much higher paragnathal count. See Charts I to IV for diagnostic characteristics.

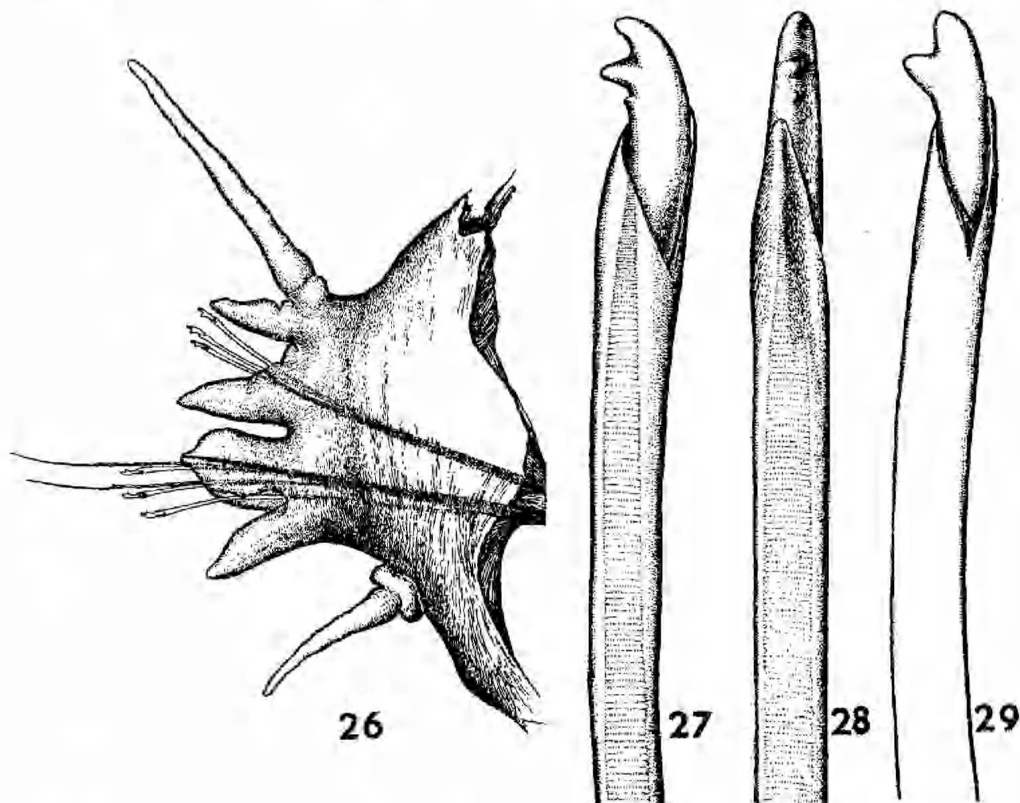


Fig. 26-29 *Nereis jacksoni*

26. Twenty-sixth parapodium seen from the front,  $\times 40$ .  
 27. A homogomph falciger from twenty-sixth parapodium seen from the side,  $\times 620$ .  
 28. A homogomph falciger from twenty-sixth parapodium seen from the cutting edge,  $\times 620$ .  
 29. A homogomph falciger from a far posterior parapodium,  $\times 400$ .

## NEREIS PERONIENSIS Kott, 1951

*Nereis callaona peroniensis* Kott, 1951, pp. 101-102, fig. 4.

This is here erected to specific category since its affinities are believed to be more remote from *Nereis callaona* Grube than its author thought. In *N. peroniensis* the homogomph falcigers of posterior notopodia taper distally to a blunt point and have a coarse tooth at the cutting edge. In *N. callaona* Grube the corresponding falciger has a much longer appendage that is distally ankylosed and there are no coarse teeth along the cutting edge (see *N. pseudonereis* Hartman 1940, pls. 26, 27, a synonym of *Nereis callaona* Grube,<sup>(2)</sup> for further characteristics). *N. callaona* Grube is known only from Peru.

<sup>(2)</sup> I am indebted to Mr. Donald J. Reish for having made a comparison of type specimens.



*N. peroniensis* Kott comes nearer *Nereis zonata persica* Fauvel, as described by Pruvot (1930, pp. 47-50, pl. 3) from New Caledonia. In both the pharyngeal armature and notopodial falcigers show great resemblance.

NEREIS COCKBURNENSIS Augener, 1913

Fig. 30-32

*Nereis cockburnensis* Augener, 1913, pp. 153-156, fig. 15 a-c.

*Localities*—Sellick Beach, S. Aust., from stones in rock pools, Apr. 1936 (about 31 individuals including some epitokes), coll. H. Hale; Sellick Beach, St. Vincent Gulf, from limestone reef covered at dead low water, Jan. 1937, (10), coll. H. M. Hale; Royal Australian Navy torpedo range at Pittwater, Broken Bay, from piles and under rocks, (5), coll. B. Dew; Pennington Bay, south coast of Kangaroo Island, (3), coll. S. J. Edmonds.

Tentacular cirri are annulate, resemble those of a eunicid (thus not as shown by Augener, 1913, pl. 3, fig. 47). The longest cirri when laid back, reach to the fifth setigerous segment.

A unique and heretofore undescribed feature is the presence of 2 kinds of notopodial falcigers. Anterior segments, from the first biramous one, have homogomph falcigers with a toothed cutting edge (fig. 30) resembling that of the corresponding neuropodial, heterogomph falciger (31). In median and posterior segments these notopodial falcigers are replaced by one which has a shorter appendage and traces of transverse ridges (fig. 32).

On the proboscis area I has a single tooth or 3 small cones in tandem (Augener described none); II has 10 cones in 2 rows (Augener gave 8 cones in 2 rows); III has only 4 to 6 cones (Augener said 2 in tandem); IV has about 18 cones in 3 rows with the largest ones on the side toward the jaws (Augener gave 5 to 11 cones in 2 or 3 rows). Area V has about 8 smaller cones or varying to only 1 cone (Augener gave 6 cones in 2 rows); VI has 5 or 4 cones in a circular area (as Augener stated); VII and VIII have a continuous band of many paragnaths with a single row of about 9 larger ones on the side toward the jaws (Augener described a broad band of many), and 5 to 7 irregular rows of many closely spaced cones on the side toward the mouth.

In postmedian segments the upper notopodial lobe comes to be small, triangular and diminishes farther back as an inconspicuous lobe.

An epitokal ovigerous individual, from Sellick Beach, 11 April 1936, has homogomph falcigers present from the first biramous parapodium. Accessory natatory lobes are first present from segment 17, at the upper base of the dorsal cirrus. Natatory setae are also present, but not yet emergent, from segment 17. The last 11 segments lack accessory lobes, indicating the presence of a third body region in epitoky.

*Nereis cockburnensis* was first described from Sharks Bay in 2-4½ meters, and Cockburn Sound, South Channel in 6½-8 metres on a rocky bottom. The present collections come from South Australia and New South Wales.

PERINEREIS Kinberg, 1866

Type P. AMBLYODONTA (Schmarda)

The 14 species indicated on Charts I to IV (see above) are largely tropical or subtropical, thus belonging mainly to the Damperian and Solanderian provinces or to New Zealand. A few species occur along southern shores of Australia (see below).

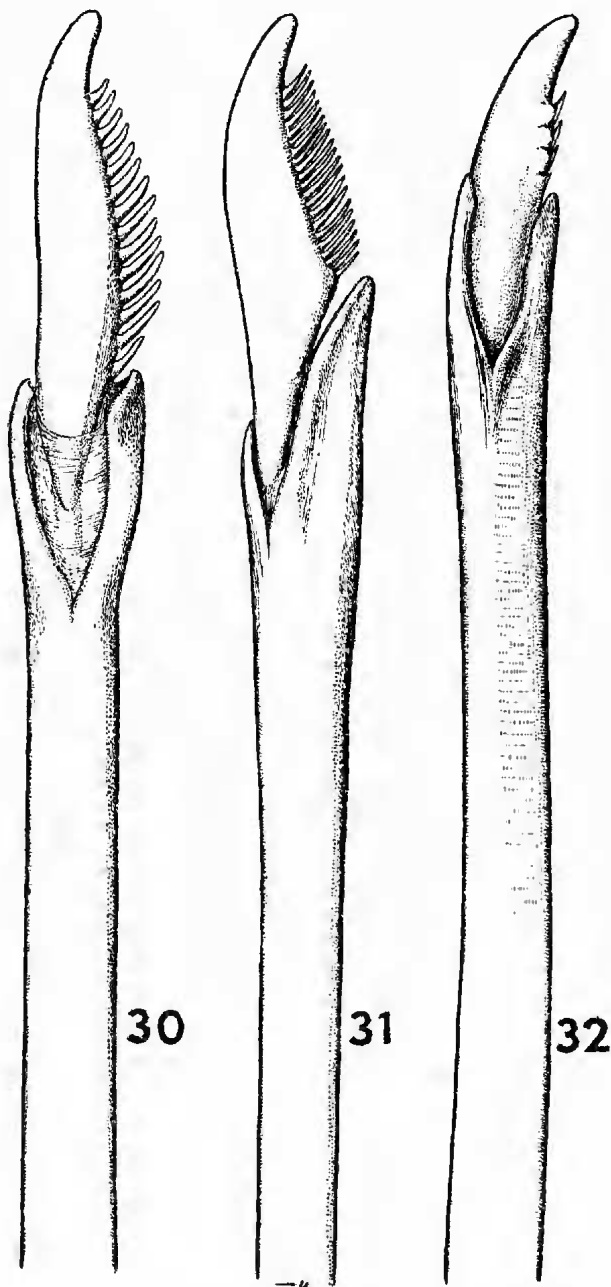
PERINEREIS AMBLYODONTA (Schmarda), 1861

*Nereis amblyodonta* Schmarda, 1861, p. 106; Ehlers, 1905, p. 28.

*Perinereis novae-hollandiae* Kinberg, 1866, p. 175 and 1910, pl. 20, fig. 9.

*Nereis (Perinereis) amblyodonta* Augener, 1913, pp. 174-175; Augener, 1922, pp. 22-23.

*Localities*—American River and lagoons, Kangaroo Island, under rocks and in mud flats; also in Venus Bay jetty, with colonies of *Galeolaria*, (8), coll. S. J. Edmonds; Port Willunga, S. Aust., in colonies of *Hormosira*, (4), coll. Miss P. Mawson; Port Jackson, Sydney Harbour, N.S.W., under rocks and in clumps of *Galeolaria*, (7), coll. B. Dew; Milsons Point, Port Jackson, N.S.W., wharf piles and on mooring chains, (1), coll. B. Dew.



*Nereis cockburnensis*

Fig. 30-32

30. A notopodial falciger from sixth parapodium, x 650.
31. A neuropodial falciger from sixth parapodium, x 650.
32. A notopodial falciger from a far posterior segment, x 650.

This species is easily identified for the presence of long dorsal lobes of posterior notopodia, and for the arrangement of paragnaths on areas V and VI. See Charts I to IV for further details and more extended distribution.

## PERINEREIS VARIODENTATA Augener, 1913

*Nereis (Perinereis) variodentata* Augener, 1913, pp. 179-182, pl. 3, fig. 50, text fig. 19.

*Localities*—Sellick Beach, St. Vincent Gulf, S. Aust., from stones in rock pools at low tide and from limestone reef covered at dead low water, (2), coll. H. M. Hale; Pt. Wynward, north-west Tasmania, April 1936, (2), coll. N. B. Tindale.

See Charts I to IV for distinguishing characteristics and distribution.

## PERINEREIS VALLATA (Grube) 1857

*Nereilepas pacifica* Schmarda, 1861, p. 107.

*Nereis (Perinereis) vallata* Augener, 1913, pp. 175-177; Augener, 1923, pp. 26-27.

*Perinereis vallata* Fauval, 1932, pp. 108-109; Knox, 1951, pp. 218-219, pls. 45-46.

*Locality*—Port Willunga, S. Aust., (1), coll. S. J. Edmonds.

See Charts I to IV for further details.

## PLATYNEREIS Kinberg, 1866

## Type P. MAGALHAENSIS Kinberg

Among the 5 species to be encountered in littoral zones in Australia and New Zealand, 1, *P. australis* (Schmarda) is perhaps limited to New Zealand; another, *P. polyscalma* Chamberlin, is tropical. *P. magalhaensis* Kinberg, *P. dumerilii antipoda*, new subspecies, and *P. bicanaliculata* (Baird) occur in the Peronian and Flindersian provinces but all are not limited to them. See also Charts I-IV, above.

## PLATYNEREIS DUMERILII ANTIPODA, new subspecies

Fig. 33-37

*Nereis (Platynereis) australis* Augener, 1913, pp. 182-184, and Augener, 1923, pp. 35-39. *Not* Schmarda, 1861.

*Localities*—Pennington Bay, south coast of Kangaroo Island, among algae, (5), coll. S. J. Edmonds; Pittwater, Broken Bay, N.S.W., on piles. Sept. 1949, associated with *Nereis cockburnensis* and *Perinereis calmani*, (2, including 1 subepitoke), coll. B. Dew; Hungry Point, Cronulla, N.S.W., Aug. 1950, (2) coll. B. Dew; Elizabeth Bay, Port Jackson, N.S.W., from mooring buoy, 28 Oct. 1950 (1 subepitoke female), coll. B. Dew; Point Wynward, north-west Tasmania, (5), coll. H. M. Hale; Sellick Beach, St. Vincent Gulf, S. Aust., limestone reef covered at dead low water (8 juveniles), coll. H. M. Hale.

These individuals have been compared with *Platynereis dumerilii* (Audouin and M. Edwards) from the Mediterranean Sea. A comparison of diagnostic parts follows:

	<i>P. d. antipoda</i> South Australia	<i>P. dumerilii</i> Mediterranean Sea
Notopodial spinigers in median and posterior segments have a length/width ratio of:	1.3/40, thus are shorter-appendaged	1/80, thus are longer-appendaged
Median parapodia, at about segment 40, have a supra-acicular fascicle of:	3 spinigers and 3 falcigers	10 spinigers only
First presence of notopodial (fig. 37) falcigers is in:	Anteromedian segments, where they are numerous and conspicuous	Postmedian segments, and they are inconspicuous and few
Acicula are coloured:	light to dark brown	black

Dorsal lobe of median and posterior segments is:	subquadrate	subtriangular
Paragnaths of area VI are:	obscure, with 2 weakly developed rows of pectinae	2 well-developed rows of pectinae
Dorsal cirri of posterior segments are:	very long (fig. 36)	not so long
In female epitoke, the upper base of the ventral cirrus has:	a long digitate lobe (fig. 35)	a short foliaceous lobe
In female epitoke, the posterior neuroacicular lobe has:	a digitate process (fig. 36)	no digitate process
In female epitoke, the parapodial change is at:	segment 22/23	segment 22-23/24 or 22/23, or 24/25

In the female epitoke the first 7 pairs of dorsal cirri enlarge (fig. 33) gradually.

The pharyngeal armature (specimen from Pt. Wynyard, Tasmania) shows areas I, II and V bare; III has 2 rows of obscure pectinae; IV is the most conspicuous area of the pharynx, with about 4 transverse series of pectinae; VI has 2 or 3 short lines of very weak pectinae; VII and VIII is an interrupted band with about 5 patches of 2 short rows each.

Individuals from Tasmania have simple gregarines in the alimentary tract, through middle and posterior third regions of the body.

*P. dumerilii ocellata* Pruvot (1930) from New Caledonia differs from *P. dumerilii antipoda* in that area VI of the proboscis is bare; the prostomium is marked with 3 dark spots, resembling eyes, hence the varietal name.

*P. dumerilii antipoda* is known only from the Flindersian province.

#### PLATYNEREIS MAGALHAENSIS Kinberg

*Platynereis magalhaensis* Kinberg, 1866, p. 177.

*Nereis (Perinereis) magalhaensis* Augener, 1923, pp. 28-39.

*Locality*—Sellick Beach, S. Aust., low tide, 16 Jan., 1936, (2), coll. H. M. Hale and K. Sheard.

This is hardly separable from *P. australis* (Schmarda) from New Zealand, except in its epitokal stages. In this the male epitoke has the first 21 segments unmodified and natatory setae from segment 22; the female has 25 segments unmodified and natatory setae from segment 26. In both species the notopodia nearly or quite lack falcigers; a weakly developed one may be found in posterior segments. Other characteristics are detailed in Charts I to IV, above.

#### PLATYNEREIS BICANALICULATA (Baird), 1863

Fig. 38, 39

*Nereis bicanaliculata* Baird, 1863, p. 109.

*Nereis agassizi* Ehlers, 1868, pp. 542-546, pl. 23, fig. 1.

*Localities*—Hungry Point, Cronulla, N.S.W., on fouling plate, 28 Sept., 1950 (1 female); Athol Bight, public jetty, 12 Oct. 1950, on kelp root (9); Camp Cove, Watsons Bay, Port Jackson, in 6-8 fms., 6 Oct. 1950 (6, including subepitokes); Port Jackson, on piles and mooring chains, 23 Oct. 1950 (15); all collections are from New South Wales, made by Miss Barbara Dew.

This strikingly characterized species is well known from the north-east Pacific as *Platynereis agassizi* (Ehlers). It was arresting to find it well represented in the collections from New South Wales. This led to a re-examination of large series from various parts of the Pacific, including some from Hawaii,

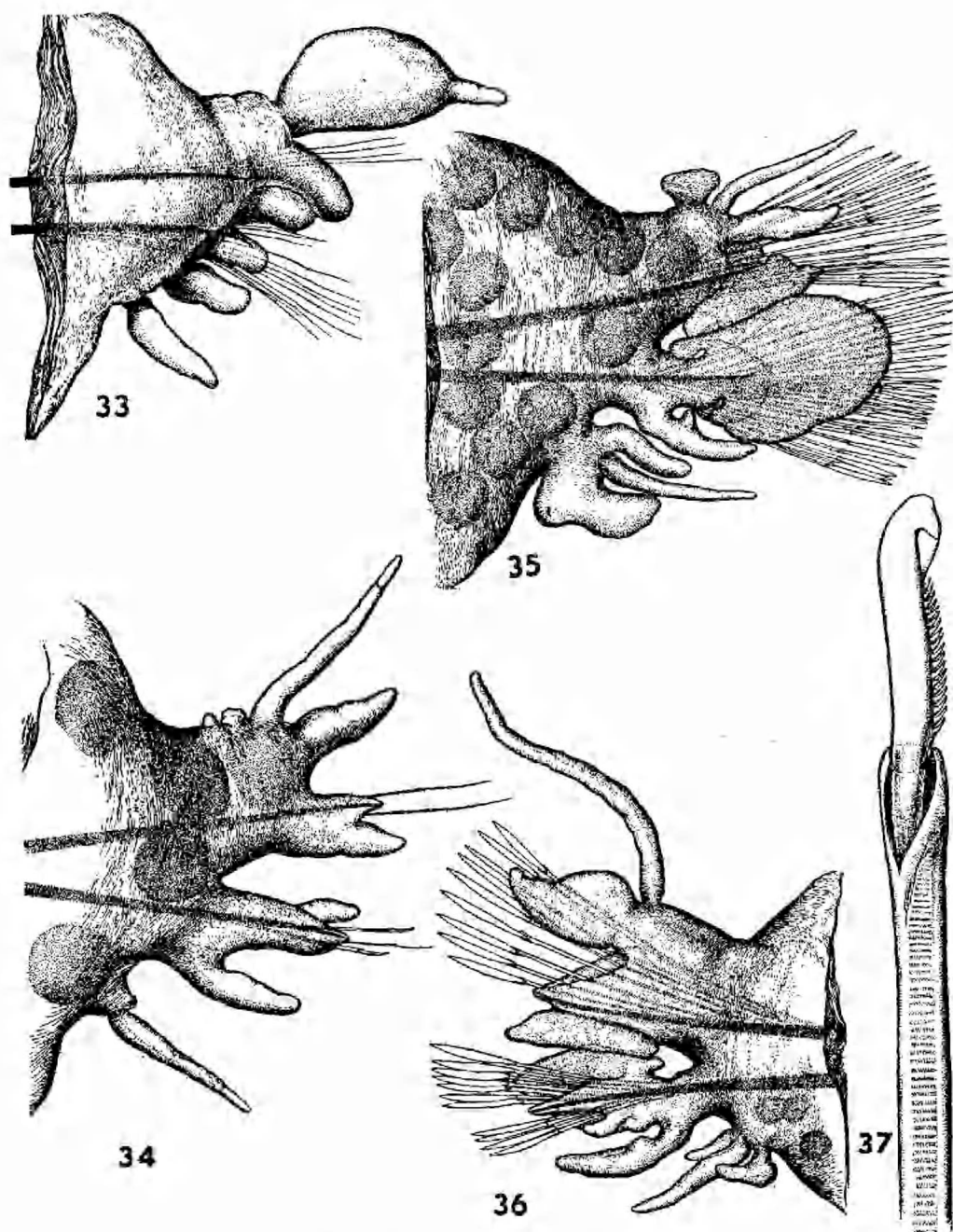


Fig. 33-37. *Platynereis dumerilii antipoda*

33. Fourth parapodium from female epitoke, showing enlarged dorsal cirrus, x 100.  
 34. Twenty-second parapodium from female epitoke, in posterior view, x 70.  
 35. Twelfth epitokal parapodium from female, in posterior view, x 50.  
 36. A far posterior parapodium from female epitoke, seen from the front, x 83.  
 37. A notopodial falciger from a posterior segment (specimen from Tasmania), x 832.

*Nereis bicanaliculata* Baird from Vancouver Island, western Canada, and many other collections from widely scattered parts of the eastern Pacific. I am unable to distinguish them morphologically, and am therefore indicating the synonymy above.

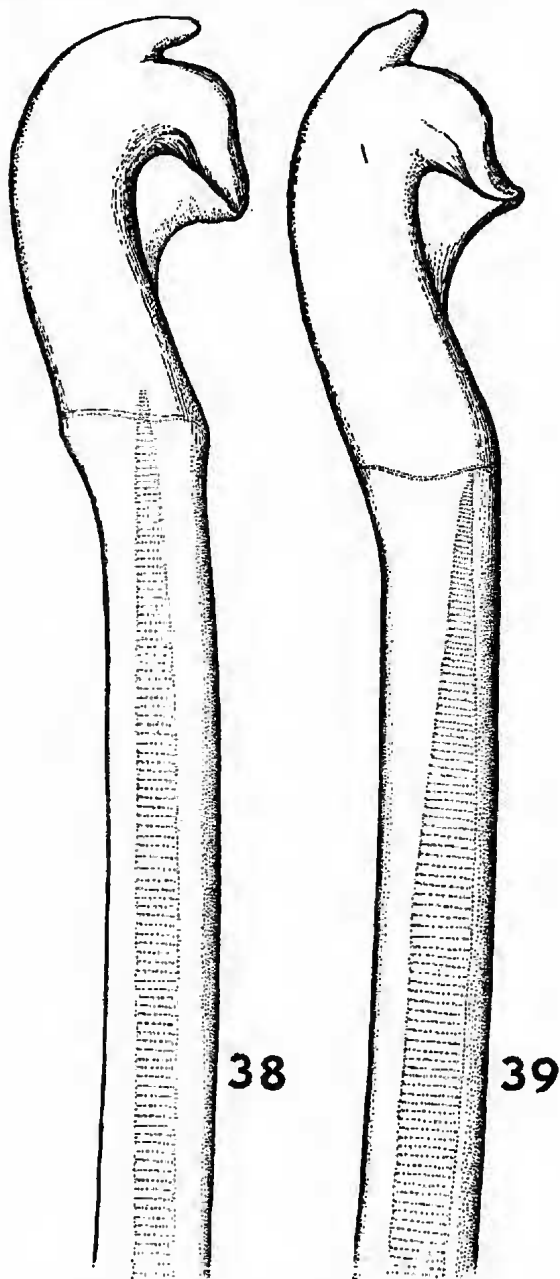


Fig. 38-39

*Platynereis bicanaliculata*

38. A simple notopodial falciger from a specimen from New South Wales, x 500.
39. A comparable notopodial falciger from one from California, x 500.

Most individuals (preserved) from New South Wales are melanistic, have paired dark patches over the sides of the body and along the parapodial bases. Large, simple, notopodial falcigers (fig. 38) are present from about segment 10 or 12 to the posterior end of the body; they occur singly or by twos or threes and have a dark brown to black tip. A corresponding falciger taken from a specimen from California is shown in fig. 39.

Neuropodial falcigers are composite, first present from about segment 50 and continue to the end of the body; they are most numerous in a fascicle in front and diminish in number behind.

On the pharynx the Australian individuals differ slightly from those of California, in that area IV of the proboscis is less, instead of more, conspicuous than area III.

The presence of simple notopodial falcigers is not limited to this genus or species, hence cannot be regarded as generic. *Ceratonereis erythraeensis* Fauvel has similar hooks in neuropodia.

The type collection of *Nereis bicanaliculata* Baird (1863) deposited in the British Museum (Natural History) contains 8 pale (faded) specimens in good condition. The largest one, somewhat over 50 mm. long (thus about 2 inches as Baird stated) consists of about 96 segments; it is posteriorly incomplete. In some individuals the parapodia are subepitokous but none has natatory setae. In all, there are one or 2 dark brown, simple notopodial falcigers, first present in parapodia from segment 12 or 13 to the posterior end. On the proboscis areas I, II and V are bare; III has a broad, oval patch; IV has a broad crescent of 7 to 10 irregular rows (this is the most conspicuous region); VI has 2 or 3 rows of pectinae; VII and VIII, continuous, have 5 transverse rows of pectinae with faint indications of 2 other rows at the ends proximal to area VI. Jaws have 7 oblique teeth and a distal fang. These individuals are inseparable from what has usually been called *Platynereis agassizi* (Ehlers), widely known from the north-east Pacific.

Throughout its range, *Platynereis bicanaliculata* is apt to occur with (or near) *Platynereis dumerilii* (Audouin and M. Edwards) or one of its varieties. They are easily separable in that *P. bicanaliculata* has large, simple falcigers in notopodia whereas *P. dumerilii* (and its subspecies or varieties) have composite falcigers in notopodia. *P. bicanaliculata* remains unknown except in northern and southern parts of the Pacific; *P. dumerilii* is cosmopolitan in warm seas.

#### LITERATURE CITED

- AUGENER, H. 1913 Die Fauna Südwest-Australiens. Polychaeta Errantia. Herausg. von Michaelsen und Hartmeyer, Jena, Bd. 4, pp. 65-304, 2 pls., 42 figs.
- AUGENER, H. 1918 Polychaeta. Beiträge zur Kenntnis des Meeresfauna West-Afrikas. Herausg. von W. Michaelsen, Hamburg, 2, Lief. 2, pp. 67-625, 6 pls.
- AUGENER, H. 1922a Litorale Polychaeten von Juan Fernandez. The Natural History of Juan Fernandez and Easter Island. Ed. by Dr. Carl Skottsberg, 3, pp. 161-218, 1 pl.
- AUGENER, H. 1922b Australische Polychaeten des Hamburger zoologischen Museums. Arch. Naturg., Berlin, 88, Abt. A, pp. 1-37, 9 figs.
- AUGENER, H. 1922c Revision der Australischen Polychaeten-Typen von Kinberg. Ark. Zool. Stockholm, 14, pp. 1-42
- AUGENER, H. 1922d Results of Dr. E. Mjöberg's Swedish Scientific Expedition to Australia, 1910-13. Polychaeten. Vetensk. Akad. Stockholm, Handl., 63, No. 6, pp. 1-49, 10 figs.
- AUGENER, H. 1923 Polychaeten von den Auckland und Campbell Inseln. Vidensk. Medd. Naturh. Foren. Copenhagen, 75, pp. 1-115, 44 figs.
- AUGENER, H. 1924 Polychaeta von Neuseeland. I. Errantia. Ibid., 75, pp. 241-441, 11 figs.
- AUGENER, H. 1927 Polychaeten von Südost- und Süd-Australien (Papers from Dr. Th. Mortensen's Pacific Expedition 1914-16. No. 38). Ibid., 83, pp. 71-275, 17 figs.
- BAIRD, W. 1863 Descriptions of several new Species of Worms belonging to the Annelida Errantia and Sedentaria or Tubicola of Milne Edwards. Zool. Soc. London, Proc., pp. 106-110
- BENHAM, W. B. 1909 Report on the Polychaeta of the sub-Antarctic Islands of New Zealand. In Reports based on observations and collections made during an expedition on the Government steamer *Hinemoa* in November 1907, 1, pp. 236-250, 1 pl.
- BENHAM, W. B. 1916a Notes on New Zealand Polychaeta. II. New Zealand Inst. Proc. Trans., 48, pp. 386-396
- BENHAM, W. B. 1916b Report on the Polychaeta obtained by the F. I. S. *Endeavour* on the coasts of New South Wales, Victoria, Tasmania and South Australia. Pt. 2. Sydney, H. C. Dannevig, 4, pls. 2 and 3, pp. 125-162, pls. 46-48

- BENHAM, W. B. 1927 Polychaeta, British Antarctic Terra Nova Expedition, 1910. Nat. Hist. Rep., Zool., 7, No. 2, pp. 47-182, 6 pls.
- BERKELEY, E., and BERKELEY, C. 1948 Annelida, Polychaeta errantia. Canad. Pac. Fauna, No. 96, (1). Fish. Res. Bd. Canada, Toronto, pp. 1-100, 160 figs.
- BLANCHARD, E. 1849 Anélides du Chili, in Gay's Historia física y política de Chile. Segun documentos adquiridos en esta republica durante doce años de residencia en ella. Zoologia. Paris, 3, pp. 9-52
- CHAMBERLIN, R. V. 1919 The Annelida Polychaeta. Mus. Comp. Zool. Harvard, Mem., 48, pp. 1-514, 80 pls.
- CLARK, H. L. 1946 The Echinoderm Fauna of Australia. Its composition and its origin. Pub. Carnegie Inst. Wash., No. 566. Washington, D.C., pp. iv and 567
- EHLERS, E. 1868 Die Borstenwürmer. Leipzig, W. Engelmann, xx and 748 pp., 24 pls.
- EHLERS, E. 1897 Polychaeten. Hamburger Magalhaenischen Sammlreise. Hamburg. Friedrichsen & Co. 148 pp., 9 pls.
- EHLERS, E. 1905 Neuseeländische Anneliden. K. Ges. Wiss., Göttingen, Math.-Phys. Kl., neue Folge, 3, No. 1, pp. 1-80, 9 pls.
- FAUVEL, P. 1911 Annélides polychètes du golfe Persique recueillies par M. M. Bogoyaw-lensky. Arch. zool. exp. gén. Paris, sér. 5, 6, pp. 353-439, 3 pls.
- FAUVEL, P. 1916 Annélides polychètes des Iles Falkland recueillies par M. Rupert Vallentin (1902-1910). Ibid., 55, pp. 417-482, 2 pls.
- FAUVEL, P. 1917 Annélides polychètes de l'Australie meridionale. Ibid., 56, pp. 159-278, 6 pls., 2 charts
- FAUVEL, P. 1918 Annélides polychètes nouvelles de l'Afrique orientale. Mus. Hist. nat. Paris, Bull., 24, pp. 503-509, 4 figs.
- FAUVEL, P. 1919 Annélides polychètes de Madagascar, de Djibouti et du golfe Persique. Arch. zool. exp. gén. Paris, 58, pp. 315-473, pls. 15-17
- FAUVEL, P. 1921 Polychètes de Madagascar, du Museum d'Histoire naturelle recueillies par M. le Dr. W. Kaudern en 1912. Ark. Zool. Stockholm, 13, No. 24, pp. 1-32, 1 pl., 2 figs.
- FAUVEL, P. 1922 Annélides polychètes de l'archipel Houtman Abrolhol (Australie occi-dentale) recueillies par M. le Professeur W. G. Dakin. Linn Soc. London, Jour., 34, pp. 487-500, 2 figs.
- FAUVEL, P. 1923 Annélides polychètes des îles Gambier et de la Guyane française. Acad. Nuovi Lincei Rome, Mem., sér. 2, 6, pp. 1-59, 8 figs.
- FAUVEL, P. 1930 Annelida Polychaeta of the Madras Government Museum. Madras Govt. Mus. Bull., n.s., Nat. Hist. Soc., 1, No. 2, pp. 1-72, 18 figs.
- FAUVEL, P. 1932 Annelida polychaeta of the Indian Museum, Calcutta. Indian Mus., Calcutta, Mem., 12, No. 1, pp. 1-262, 9 pls., 40 figs.
- FAUVEL, P. 1933 Annélides polychètes du golfe du Pei Tchen Lyde la collection de Musée Hoang ho Pai ho. Mus. Hoangho Pai ho Tien-Tsin, Pub., 15, pp. 1-67, 6 figs.
- FAUVEL, P. 1936 Sur quelques Annélides polychètes de l'île de Paques. Mus. Hist. nat. Paris, Bull., sér. 2, 8, pp. 257-258
- FAUVEL, P. 1947 Annélides polychètes de Nouvelle-Calédonie et des îles Gambier. Faune de l'Empire Français. VIII. 108 pp., 90 figs.
- FEUERBORN, H. 1931 Neue marine Einwanderer der Binnengewässer von Java und Sumatra. Forschung und Fortschritte, Berlin, No. 17
- GOODRICH, E. 1945 The study of nephridia and genital ducts since 1895. Micr. Sci. London, Quar. Jour., 85, \*2-4, pp. 113-392, figs.
- GRAVELY, F. 1927 The littoral fauna of Krusadai Island in the Gulf of Mannar. Chaetopoda. Madras Govt. Mus., Bull., n.s., Nat. Hist., 1, pp. 55-86, 2 pls.
- GRAVIER, C. 1901 Contribution à l'étude des Annélides polychètes de la Mer Rouge. Nouv. Arch. Mus. Paris, sér. 4, 3, pp. 147-268, figs. 160-285, 4 pls.
- GRUBE, E. 1857 Annulata Oerstediana. Pt. 2, pp. 158-166
- GRUBE, E. 1868 Reise der Oesterreichischen Fregatte *Novara* um die Erde in den Jahren 1857, 1858 und 1859. Zool. Theil, Bd. 2, Abt. 3. Anneliden. Wien pp. 1-48, 4 pls.
- GRUBE, E. 1869 Beschreibung neuer von der *Novara* Expedition mitgebrachter Anneliden und einer Landplanarie. Zool.-Bot. Gesells., Wien, Verh., 16, pp. 173-184
- GRUBE, E. 1878 Annulata Semperiana. Acad. Sci. St. Petersburg, Mem., 25, 300 pp., 15 pls.
- HARTMAN, O. 1940 Polychaetous annelids. Pt. 2. Chrysopetalidae to Gomaididae. Hancock Pacific Exped., 7, pp. 173-287, 14 pls.
- HARTMAN, O. 1948 The Marine Annelids erected by Kinberg with notes on some other types in the Swedish State Museum. Ark. Zool. Stockholm, 42A, pp. 1-137, 18 pls.
- HARTMAN, O. 1952 *Iphitime* and *Ceratocephala* (Polychaetous annelids) from California. Southern Calif. Acad. Sci., Bull., 51, pp. 9-20, 2 pls.
- HARTMAN, O. 1953 Non-pelagic Polychaeta. Swedish Antarctica Exped. 1901-1903. Further Zoological Results of the Swedish Antarctic Expedition, 4, No. 11, pp. 1-83, 1 chart, 21 figs.



- HORST, R. 1889 Contributions towards the Knowledge of the Annelida Polychaeta. Notes Leyden Mus. Jentink, 11, pp. 161-186, 2 pls.
- HORST, R. 1924 Polychaeta errantia of the *Siboga*-Expedition. Nereidae and Hesionidae. *Siboga*-Exped. Leyden, 99, (Monogr. 24, 1c), pp. 145-198, 7 pls.
- KINBERG, J. 1866 Annulata nova. Oerv. Vet. Akad. Stockholm, 22, pp. 167-169 and 239-258
- KINBERG, J. 1910 Annulater. Kongliga Svenska Fregatten Eugénies Resa omkring jorden, 1851-53. Uppsala and Stockholm, Almqvist and Wicksells., 78 pp., 29 pls.
- KNOX, G. A. 1951 The Polychaetous Annelids of Banks Peninsula. Rec. Canterbury Mus., 5, pp. 213-229, pls. 44-50
- KOTT, PATRICIA 1951 Nereidae and Eunicidae of South-western Australia; also notes on the Ecology of Western Australian Limestone Reefs. Roy. Soc. West. Australia, Jour., 35, pp. 85-130, 7 figs.
- McINTOSH, W. C. 1885 Report on the Annelida Polychaeta collected by H.M.S. *Challenger* during the years 1873-76. *Challenger* Reports, 12, pp. 1-554, 55 and 39a pls.
- MARENZELLER, E. VON. 1879 Südjapanische Anneliden. Akas. Wiss. Wien, Denkschr., 41, pp. 109-152, 6 pls.
- MONRO, C. 1926 On the Polychaeta collected by H.M.S. *Alert*, 1881-1882. Families Hesionidae and Nereidae. Linn Soc. London, Jour., 36, pp. 311-323
- MONRO, C. 1930 Polychaete Worms. *Discovery* Reports, 2, pp. 1-222, 91 figs.
- MONRO, C. 1931 Polychaeta, Oligochaeta, Echiuroidea and Sipunculoidea. Great Barrier Reef (Qld.) Exp. 1928-29. Sci. Rep. Brit. Mus. (Nat. Hist.), 4, (1), pp. 1-37, 15 figs.
- MONRO, C. 1936 Polychaete Worms. II. *Discovery* Reports, 12, pp. 59-198, 34 figs.
- MONRO, C. 1937 The John Murray Expedition 1933-34. Scientific Reports, Polychaeta. 8, No. 8, pp. 243-321, 28 figs.
- MONRO, C. 1938 On a small collection of Polychaeta from Swan River, Western Australia. Ann. Mag. Nat. Hist. London, ser. 11, 2, pp. 614-624, 13 figs.
- MONRO, C. 1939a Polychaeta Antarctic Research Expedition, 1929-1931. Adelaide, Australia. Reports, Ser. B (Zoology and Botany), 4, pt. 4, pp. 89-156, 28 figs.
- MONRO, C. 1939b On some Tropical Polychaeta in the British Museum, mostly collected by Dr. C. Crossland at Zanzibar, Tahiti and the Marquesas. Novitat. Zool. London, 41, pp. 302-305
- OKUDA, S. 1938 Polychaetous Annelids from the vicinity of the Mitsui Institute of Marine Biology. Japan. Jour. Zool., 8, pp. 75-105, 15 figs.
- OKUDA, S. 1940 Polychaetous Annelids of the Ryukyu Islands. Biogeogr. Soc. Japan, Bull., 10, No. 1, pp. 1-24, 9 figs.
- PRUVOT, G. 1930 Annélides Polychètes de Nouvelle-Calédonie recueillies par M. François. Arch. zool. exp. gen. Paris, 70, pp. 1-94, 8 figs., 3 pls.
- QUATREFAGES, A. DE. 1865 Histoire naturelle des Années marina et d'eau douce. Paris. 1, pp. 1-588
- RACOVITZA, E. 1893 Sur la *Micronereis variegata* Clpd. Acad. Sci. Paris, C.R., 116, pp. 1,390-1,392
- SCHMARDT, L. 1861 Neue wirbellose Thiere beobachtet und gesammelt auf einer Reise um die Erde, 1853 bis 1857. Leipzig. Pt. 2. pp. 1-164, 22 pls.
- WILLEY, A. 1905 Report on the Polychaeta collected by Professor Herdman, at Ceylon, in 1902. Ceylon Pearl Oyster Fisheries, Suppl. Rep., pt. 4, pp. 243-324, 8 pls.